

Effect of Pre-Sowing Treatments with Uv-Ray on the Chemical Components of Fennel (*Foeniculum Vulgare Mill*)

Ashti Sleman Abdulla¹ and Ahmad H. Ameen H. Rashid¹⁺

¹Field Crops Dep., Faculty of Agricultural Sciences, University of Sulaimani, Kurdistan Region - Iraq.

Abstract. The objective of the present study was to evaluate the effect of pre-sowing treatments with UV-ray on the amount of essential oil and its components. The results showed that exposing seeds to UV-A \times D₁ \times T₁ significantly increased protein content comparing to control. Meanwhile it decreased the concentration of *trans*-anethole. However increasing time to 40 min. (UV-A \times D₁ \times T₂) decreased protein content. Concerning the studied chemical compounds, they were significantly affected by UV-ray, but each had an optimum dose for accumulating a high rate. When seeds were exposed to UV-C \times D₂ \times T₂ produced the higher concentrations of *trans*-anethole as the main compound of fennel essential oil and M-chavicol (estragol), compared to control. And (UVC \times D₁ \times T₃) also increased qualitative characters such as essential oil content, essential oil yield and fixed oil content compared to control. Protein profiles showed around 9 protein bands, clearly dominant. 1, 2 and 3 protein bands with down regulations were identified when seeds exposed to UV-A \times D₂ \times T₂ (mutant M₂) and UV-C \times D₁ \times T₃ (M₁₂). This was conformed the under production of cellular protein by UV mutation. SDS-polyacrylamide gel electrophoresis (PAGE) analysis did not show additional or deletion in number of protein bands in UV mutants.

Keywords: *Foeniculum vulgare Mill*; Protein content; Trans-anethole; UV-ray.

1. Introduction

Fennel (*Foeniculum vulgare Mill.*) which belongs to the family Apiaceae, was selected. Fennel is one of the aromatic and medicinal plants, small group of annual, biennial or perennial herbs [1]. Fennel is used as a spice and also as an important ingredient in various folklore medicines throughout the world [2].

Ultraviolet radiation is the part of the non-ionizing region of the electromagnetic spectrum that comprises approximately 8-9 % of total solar radiation [3]. UV-C (100-280 nm) is extremely active photochemically and biologically lethal, but it is completely excluded by the ozone layer. UV-B region (280- 320 nm) is the part of ultraviolet radiation which is biologically active and vulnerable to significant changes resulting from differences in the extent of blocking by the ozone layer. UV-A (320-400 nm) is present in significant amounts in natural daylight and is relatively little affected by changes in the ozone layer [4].

In recent years, there has been an increased interest in UV light techniques, because ultraviolet radiation is one important factor that in many cases has an effect on plant growth and stimulates the production of secondary metabolites. The degree of the effect of UV-ray on plant growth varies according to plant species, but their effecting on plant secondary metabolites may return to some specific reasons such as the secondary metabolites which have a sun screening effect and protect the cells from the radiation, this would be especially likely if the metabolites are concentrated in the epidermis or other superficial tissues, and the radiation aids in reaching a certain level of general differentiation necessary for production of secondary metabolites[5].

⁺ Corresponding author. Tel.: + 0964 533370095.
E-mail address: drahamad1955@gmail.com.

2. Materials and Methods

This experiment was carried out at the research station field of the Faculty of Agricultural Sciences / Sulaymani University in Bakrajo during the season (October 2011- July 2012). The seeds of *Foeniculum vulgare* Mill. were (local variety) standard seeds obtained from Pakan Bazaar product of Iran and before sowing they were treated with ultraviolet ray {Control: UV-C: 254 nm and UV-A: 366 nm} at two different distances (D_1 :6 cm and D_2 : 12 cm) between seed lots and the source of UV-ray for different periods of time (T_1 : 20, T_2 : 40, and T_3 : 60 min.). The field land was designed as a factorial experiment conducted in split-split plot design, with three replicates.

2.1. Studied Characters

2.1.1. Chemical components

2.1.1.1. Protein %

Protein content was determined in seeds by using Kjeldahl method [6] and for calculation, the following equations were used:

- $T. N\% = (V \times N \times E \times 0.014 / S) \times 100$
- $Protein\% = T.N\% \times 6.25$
- T.N= Total nitrogen
- V= Volume of 0.01 N HCL titrated for the sample (ml).
- N = Normality of HCL solution.
- E= Equivalent weight of nitrogen

2.1.1.2. Fixed oil %

The ratio (%) of fixed oil was determined by using the Soxhlet apparatus, the method used by [7], then fixed oil content calculated as follows [8]:

- $Fixed\ oil\% = (Weight\ of\ flask\ after\ extraction - weight\ of\ flask\ prior\ to\ extraction / Weight\ of\ sample) \times 100$

2.1.1.3. Essential oil %

Water steam distillation: Essential oil was quantified by using a Clevenger apparatus, according to [9]. Samples of 25 g from the dried and cleaned mixed seeds for each treatment were ground up; therefore, they were subjected to hydro-distillation for 120 min. [10] to obtain the yield of essential oil and the chemical component of the oil samples, using High Performance Liquid Chromatography (HPLC) technique. The following equation was used to determine the content of essential oil.

- $Essential\ oil\% = Volume\ of\ essential\ oil / Weight\ of\ sample \times 100$

And essential oil yield was calculated from the following formula:

- $Essential\ oil\ yield\ kg\ h^{-1} = Essential\ oil\% \ w/w \times seed\ yield\ kg\ h^{-1}$

2.1.1.4. Essential oil analyzing

The extracts (essential oil) were qualitatively and quantitatively analyzed by High Performance Liquid Chromatography (HPLC) [Research Laboratory of the Green Field Company in Baghdad] using a model Shimadzu Corporation, Kyoto Japan, LC- 10 AV equipped with binary delivery pump model LC-10A Shimadzu. Column: Phenomenex C-18, 3 μ m particle size (50 \times 4.6 mm I.D.) [11].

Calculation: Concentration of sample μ m/ml = (Area of sample/ Area of standard) \times Concentration of standard \times dilution factor [12].

2.1.1.5. Protein extraction

Protein extracted in seeds by using gel electrophoresis [13].

2.2. Statistical Analysis

Statistical analysis of the data was carried out by using statistical program JMP Stat, for checking the significance of the different treatments, whereas LSD at 5% probability levels ($P < 0.05$) was used to compare the differences among the treatments.

3. Results and Discation

The results of (Table 1) indicated that irradiation of seeds to UV had significant effect on qualitative characters of fennel seeds. Exposing seeds to UV for T3 (60 min.) gave significantly the highest value of essential oil percent accumulated in seeds (3.9 %), followed by T2 (40 min.) with (3.6%). Similar results with positive effect of UV-ray on essential oil concentration in anise were found by [14], in Parsley by [15], and in lavender by [16].

Table 1. Effect of exposure time to UV ray on essential oil content (EO %), essential oil yield (EOY kg ha⁻¹, fixed oil content (FO %) and protein content (P %) in fennel seeds.

Character	Exposure time			LSD 0.05
	T ₁	T ₂	T ₃	
EO%	3.4	3.6	3.9	0.11
EOY(Kg h ⁻¹)	51.7	62.6	55.9	2.8
FO%	6.6	19.5	19.7	0.8
P%	20.5	19.5	19.7	0.8

T₁:20 min., T₂:40 min., T₃:60 min.

Table 2 verifies the interaction between UV-C×D₁×T₃ produced higher values of essential oil content (4.5%) and essential oil yield (82.4 kg ha⁻¹) compared to control (3.6 % and 45.9 kg ha⁻¹), respectively, this interaction increased essential oil concentration by (25%), and essential oil yield by (79.5%). And the lowest value of essential oil yield was found by UV-C×D₁×T₁ (32.8 kg ha⁻¹). The results were in agreement with those by [17], who reported an increase (50%) of essential oil production when *Mentha spicata* irradiated by UV-ray.

Interaction between UV-C×D₁×T₃ significantly increased fixed oil (7.3%) compared to control (7.1%), but the lowest value with (5.6%) was found by the interaction UVC×D₂×T₂. Irradiation of seeds with UV-A×D₁×T₁ significantly increased protein content (22.3%), followed by UV-C×D₁×T₂ (21.6%) compared to control (19.8%) where their contents were increased by (12.6%) and (9.0%), respectively. These results were in agreement with those by [18] who reported an increase in protein content of *Brassica napus* plants when exposed to UV-ray. This increase was probably due to the synthesis of defense proteins and enzymes. The results of the same table showed that the minimum value of protein content (18.2%) was produced by the interaction UV-C×D₁×T₃.

The significant interaction between UV-C×D₁×T₃ increased fixed oil% ,essential oil% and essential oil yield by (2.8%, 25 % and 79.5%), respectively and protein content was increased by (9%) with the interaction UV-C×D₁×T₂ compared to control. The interaction between UV-A×D₁×T₁ increased protein concentrations by (12.6%) compared to control.

Table 2. Effect of UV_{wL}, distance and exposing time on fixed oil rate (FO %), protein (P %), essential oil (EO %) and essential oil yield (EOY kg ha⁻¹) in fennel seeds.

Character	EO%	EOY(Kg h ⁻¹)	FO%	P%
Control	3.6	45.9	7.1	19.8
UV-C×D ₁ ×T ₁	2.9	32.8	6.1	19.9
UVC×D ₁ ×T ₂	4.0	64.0	6.0	21.6
UVC×D ₁ ×T ₃	4.5	82.4	7.3	18.2
UVC×D ₂ ×T ₁	4.0	62.8	6.3	20.5
UVC×D ₂ ×T ₂	3.2	67.7	5.6	18.2
UVC×D ₂ ×T ₃	4.0	50.8	6.5	19.9
UVA×D ₁ ×T ₁	3.3	72.3	6.0	22.3
UVA×D ₁ ×T ₂	4.0	69.9	6.3	17.9
UVA×D ₁ ×T ₃	4.2	47.6	6.8	20.9
UVA×D ₂ ×T ₁	3.3	50.7	6.8	20.6
UVA×D ₂ ×T ₂	3.6	81.9	6.1	19.6
UVA×D ₂ ×T ₃	3.7	63.0	6.8	19.5
LSD 0.05	0.2	6.9	0.8	2.0

Control, UV-C: 354 nm, UV-A: 365 nm, D₁:6cm, D₂:12 cm, T₁:20 min., T₂:40 min., T₃: 60 min.

Table 3 shows that the exposed seeds to UV-C and UV-A decreased concentrations of α -pienene, methyl chavicol, *trans*-anethole, fenchone and terpenene compared to control by (580.9%, 80%, 51.9%, 53.9% and 9.4%) compared to the lowest value for each treatment. But exposed seeds to UV-A significantly increased concentrations of eucalyptol, limonene, α - phellandrene and *p*-cymene to (1.50, 1.22, 0.93 and 0.82 mg ml⁻¹), respectively.

Exposed seeds to UV-C wavelength significantly increased concentration of estragole (methyl chavicol) and *trans*-anethole to (1.26 and 11.09 mg ml⁻¹), respectively compared to UV-A (1.06 and 8.76 mg ml⁻¹), while exposed seeds to UV-A wavelength significantly increased concentration of α -pinene, fenchone, eucalyptol, limonene, terpenene, α -phellandrene and *p*-cymene (1.05, 5.90, 1.50, 1.22, 0.86, 0.93 and 0.82 mg ml⁻¹), respectively compared to UV-C wavelength. These variations in the contents of chemical components of fennel seeds due to exposing seeds to different UV wavelengths could be explained by the different pathways through which each chemical is synthesized, such as the phenylpropanoid *trans*-anethole and estragole, found in the essential oil of fennel, are synthesized by shikimic acid pathway, while limonene, terpenene and α - pinene synthesized by acetate-mevalonic acid pathway [19].

Table 3. Effect of UV-ray (A and C) doses on chemical components of fennel essential oil as identified by (HPLC).

Chemical component	α -p mg ml ⁻¹	m.ch mg ml ⁻¹	<i>t</i> -a mg ml ⁻¹	fe mg ml ⁻¹	eu mg ml ⁻¹	Li mg ml ⁻¹	ter. mg ml ⁻¹	α - ph mg ml ⁻¹	<i>p</i> -cy mg ml ⁻¹
UV treatment									
Control	2.86	1.91	13.31	6.05	0.82	0.55	0.95	0.54	0.76
UV-C	0.42	1.26	11.09	3.93	0.94	0.57	0.56	0.44	0.11
UV-A	1.05	1.06	8.76	5.90	1.50	1.22	0.86	0.93	0.82
LSD0.05	0.012	0.002	0.014	0.011	0.019	0.015	0.023	0.11	0.04

Control, UV-C: 354 nm, UV-A: 365 nm.

The results of Table 4 showed that irradiation of fennel seeds to UV ray for 60 min. (T₃) significantly increased concentration of α -pienene, *trans*-anethole, terpenene and *p*-cymene (1.60, 12.03, 0.83 and 0.59 mg ml⁻¹), respectively. When exposed time was decreased the concentrations of these chemical components were decreased. Exposed seeds to UV-ray for 40 min. (T₂) significantly increased the concentrations of methyl chavicol, eucalyptol, limonene and α -phellandrene (1.92, 1.19, 0.84 and 0.73 mg ml⁻¹), respectively. While exposed seeds to UV-ray for 20 min. (T₁) significantly increased the concentration of fenchone (5.86 mg ml⁻¹) and with increasing exposed time, the concentration of fenchone was decreased.

Table 4. Effect of seed exposure to different times of UV-ray on chemical components of fennel essential oil as identified by HPLC.

Chemical component	α -p mg ml ⁻¹	m.ch mg ml ⁻¹	<i>t</i> -a mg ml ⁻¹	fe mg ml ⁻¹	eu mg ml ⁻¹	li mg ml ⁻¹	ter. mg ml ⁻¹	α - ph mg ml ⁻¹	<i>p</i> -cy mg ml ⁻¹
Time (min.)									
T1 (20 min.)	1.41	1.09	10.32	5.86	1.06	0.77	0.74	0.57	0.54
T2 (40 min.)	1.32	1.92	10.81	4.32	1.19	0.84	0.81	0.73	0.55
T3 (60 min.)	1.60	1.22	12.03	5.69	1.02	0.72	0.83	0.62	0.59
LSD0.05	0.011	0.001	0.006	0.009	0.007	0.007	0.009	0.08	0.011

T₁:20 min., T₂:40 min., T₃: 60 min.

The results of Table 5 revealed that exposing seeds to UV-A \times D₁ \times T₃ interaction significantly increased the concentrations of α -pinene (3.03 mg ml⁻¹) and terpenene (1.46 mg ml⁻¹) compared to control (2.86 and 0.95 mg ml⁻¹), respectively. The interaction between UV-C \times D₂ \times T₂ significantly increased the concentrations of methyl chavicol and *trans*-anethole (4.33 and 14.83 mg ml⁻¹), respectively compared to control (1.91 and 13.31 mg ml⁻¹). The results showed that after exposure of seeds to UV-C treatment increased the concentrations of compounds synthesized by shikimic acid pathway, while the concentrations of limonene, synthesized by acetate-mevalonic acid Pathway were decreased [19], this explained that UV-C had positive effect on shikimic acid pathway and negative effect on acetate-mevalonic acid pathway. The interaction between UVA \times D₂ \times T₁ significantly increased concentration of fenchone (10.67 mg ml⁻¹) compared to control and the rest of other treatments for the same compounds, while interaction between UVA \times D₁ \times T₂ gave the lowest value for the concentration of fenchone (0.78 mg ml⁻¹). The interaction between UV-A \times D₂ \times T₂ significantly increased the concentration of eucalyptol (2.25 mg ml⁻¹) compared to control and the rest of

other treatments for the same compounds, while interaction between UVA×D₂×T₃ gave the lowest concentration value of eucalyptol (0.35 mg ml⁻¹).

The interaction between UV-A×D₁×T₂ significantly increased the concentration of limonene, α-phellandrene and *p*-cymene (2.07, 2.15 and 1.45 mg ml⁻¹), respectively compared to control treatments (0.55, 0.54 and 0.76 mg ml⁻¹), respectively. The interaction between UV-C×D₂×T₁ gave the lowest concentration values of terpenene and α-phellanderene (0.18 and 0.14 mg ml⁻¹), respectively. The interaction between UV-A×D₂×T₃ gave the lowest concentration values of eucalyptol and limonene (0.35 and 0.150 mg ml⁻¹), respectively.

Table 5. Interaction effect of UV_{WL}, distance and exposure time on chemical components of fennel essential oil as identified by HPLC.

Chemical component	α-p mg ml ⁻¹	m.ch mg ml ⁻¹	t-a mg ml ⁻¹	fe mg ml ⁻¹	eu mg ml ⁻¹	li mg ml ⁻¹	ter. mg ml ⁻¹	α-ph mg ml ⁻¹	p-cy mg ml ⁻¹
UV _{WL} ×D×T									
Control	2.86	1.91	13.31	6.05	0.82	0.55	0.95	0.54	0.76
UV-C×D ₁ ×T ₁	0	0.32	5.90	6.74	1.14	1.06	0.91	0.66	0
UV-C×D ₁ ×T ₂	1.18	0	8.51	5.15	1.44	0.35	0.65	0.54	0.15
UV-C×D ₁ ×T ₃	0	0.81	14.54	3.39	1.11	0.86	0.74	0.64	0
UV-C×D ₂ ×T ₁	1.008	1.63	14.49	2.72	0.76	0.35	0.18	0.14	0.14
UV-C×D ₂ ×T ₂	0	4.33	14.83	1.25	0.44	0.45	0.34	0.27	0
UV-C×D ₂ ×T ₃	0.37	0.45	8.26	4.34	0.76	0.35	0.57	0.38	0.36
UV-A×D ₁ ×T ₁	1.44	0.58	4.49	2.93	1.55	1.27	0.52	0.85	0.75
UV-A×D ₁ ×T ₂	0	2.86	5.86	0.78	1.37	2.07	1.37	2.15	1.45
UV-A×D ₁ ×T ₃	3.03	1.85	14.04	4.52	2.23	1.85	1.46	1.16	0.94
UV-A×D ₂ ×T ₁	0.33	0.16	10.42	10.67	1.25	0.86	0.93	0.67	0.85
UV-A×D ₂ ×T ₂	1.01	0.51	9.02	6.65	2.25	1.09	0.57	0.32	0.16
UV-A×D ₂ ×T ₃	0.505	0.38	8.72	9.83	0.35	0.15	0.33	0.45	0.75
LSD0.05	0.02	0.003	0.016	0.022	0.018	0.018	0.023	0.21	0.028

Control; UV-C: 254 nm, UV-A: 365 nm D₁: 6 cm, D₂: 12 cm, T₁: 20 min., T₂: 40 min., T₃: 60 min.

Fig. 1 shows the effect of UV_{WL} (A and C) on gene expression in fennel seeds. The results revealed that the profiles of the extracted proteins from different genotypes, visualized on 15% SDS-PAGE, displayed few differences indicating the effect of the mutagen effect. Protein profiles showed that around 9 protein bands were clearly dominant. 1, 2 and 3 protein bands with down regulation were identified in the UV mutants M₂ and M₁₂, respectively. This was confirmed the under production of cellular protein by UV mutation. SDS-poly acrylamide gel electrophoresis (PAGE) analysis did not show additional or deletion in number of protein bands in UV mutants.

Earlier studies by [20] showed that the UV radiation damages ribosomes by cross linking cytosolic ribosomal proteins S14, L23a, and L32, and chloroplast ribosomal protein L29 to RNA as [21] reported the synthesis of new protein bands under UV-C stress. UV damages to specific binding proteins are considered to play an important role in early responses of cells irradiated with UV, including damage of recognition in the DNA repair process. Exposure to UV-B light produced effects which were very similar to those obtained from UV-C exposure [22]. UV-C and UV-B rays were found to cause a decline in total RNA content, enzyme activity and protein levels of several key photosynthetic proteins including RUBISCO [23] and [24] and also decline in D1 polypeptide of photosystem II, chlorophyll a/6-binding protein and the ATPase complex [25].

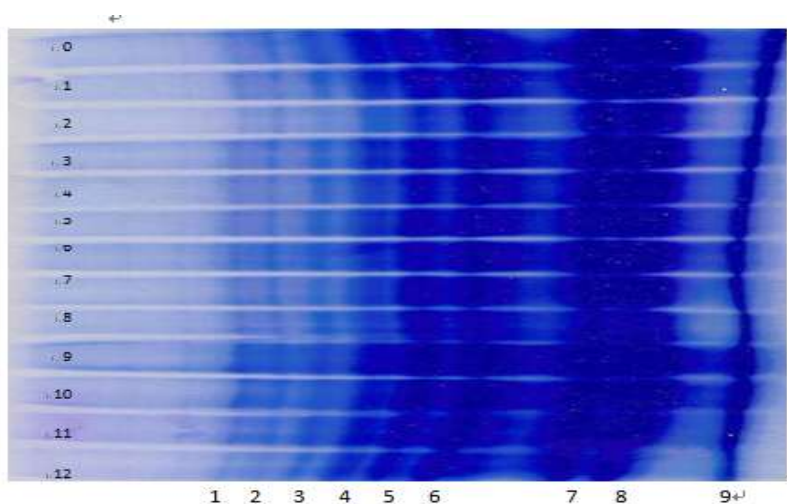


Fig. 1: Effect of UV_{WL} (A and C) on gene expression in fennel seed:

UV- 0 (Control);

M₁: UV-A × D₂ × T₁; (UV-A at distance 12 cm for 20 min.)

M₂: UV-A × D₂ × T₂; (UV-A at distance 12 cm for 40 min.)

M₃: UV-A × D₂ × T₃; (UV-A at distance 12 cm for 60 min.)

M₄: UV-A × D₁ × T₁; (UV-A at distance 6 cm for 20 min.)

M₅: UV-A × D₁ × T₂; (UV-A at distance 6 cm for 40 min.)

M₆: UV-A × D₁ × T₃; (UV-A at distance 6 cm for 60 min.)

M₇: UV-C × D₂ × T₁; (UV-C at distance 12 cm for 20 min.)

M₈: UV-C × D₂ × T₂; (UV-C at distance 12 cm for 40 min.)

M₉: UV-C × D₂ × T₃; (UV-C at distance 12 cm for 60 min.)

M₁₀: UV-C × D₁ × T₁; (UV-C at distance 6 cm for 20 min.)

M₁₁: UV-C × D₁ × T₂; (UV-C at distance 6 cm for 40 min.)

M₁₂: UV-C × D₁ × T₃; (UV-C at distance 6 cm for 60 min.)

4. References

- [1] Sharma, R. 2000. Medicinal plants of India: An Encyclopedia. Daya Jur. Publishing House, Delhi : 108-109.
- [2] WHO, (World Health Organization). 2005. Global Atlas of Traditional, complementary and Alternative Medicine: World Health Organization, Geneva.
- [3] Fredreick, J. E. 1993. Ultraviolet sunlight reaching the Earth's surface. A Review of recent research. *Photochem. Photobiol.* 57, 175-178.
- [4] Wilson, CH. L. and G. F. Heidinger, 2012. Ultraviolet light treatments for increasing seed yields. Horizon seed technologies, inc.jacksonville, IL (US). 2-11.
- [5] Zhang J. E. and L. O. B jorn. 2009. The effect of ultraviolet radiation on the accumulation of medicinal compounds in plants *FITOTE-01795*; No of Pages 12 *journal homepage: www.elsevier.com/locate/fitote*.
- [6] A O. C. A, 1995. *Official Methods of Analysis*, (16th Ed.) Arlington Va USA: Association of 269 Official Analytical Chemists.
- [7] A O. C. A, 1990 in *Official Methods of Analysis*, (15th Ed.) Association of Official Analytical Chemists, Washington, DC.
- [8] Nik, Nik, A. R.; S. Al- Rawi.; Md. S. Hossain; Ibrahim; M. A. Abdul majid, M.O. and Ab. Kadir. 2011. Malaysia. Comparative Study between the Supercritical Extraction and Soxhlet Extraction of Nutmeg Seed.
- [9] European pharmacopoeia. 2000. (3rd Ed.). Dritter Nachtrag, Council of Europ, Strasbourg: 499-500.
- [10] Ozel, A. 2008. Anise (*Pimpinella anisum* L.): change in yield and component composition on harvesting at

different stage of plant maturity. *Expl Agric. Cambridge University Press*, volume 45, 117–126.

- [11] Bisignano, G.; K. Sanogo; A. Masino; R. Aquino; U.D. Angelo; M.P. Germano; R.D. pasquale and C. Pizza. 2000. Antimicrobial activity of *Mitracarpus scaber* extracts and isolated constituents. *Phyto.chem. Letters in applied microbiology*. 30: 105-108.
- [12] Illiana, I.; L. Witte and A. W. Alfermann. 1989. Production of alkaloid by transformed root cultures of *Datura innoxia*. *Planta Medica*, Vol. 55: 229-230.
- [13] Laemmli, U. K. 1970. Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature* 227: 680-685.
- [14] Al- Akrawi, H. S. K., 2006. Exposure of seeds, Organs and Callus of *Pimpinella anisum* L. to ultraviolet rays and determination of Anethole content by high performance liquid chromatography. M.sc. Thesis. Mosul Uni. College of Education.
- [15] Al- Zaidi, R. N. J. 2005. The effects of Gamma and Ultraviolet rays in tissue culture, the content of protein, nucleic acid and volatile oils in parsley plants *Petroselinum crispum* L. Musil university. M.sc. Thesis. College of Education.
- [16] Badawy, E.; S. Sakr; M. El- Sharnouby; E. Szoke; I. Mathe; G. Blunden, A. Kery. 2003. Production and composition of Lavender plants through tissue culture as affected with gamma irradiation treatments. *Actahorticulture*. 597: 325-328.
- [17] Karousou, R.; G. Grammatikopoulos; T. Lanaras; Y. Manetas and Kokkin *phytochemistry*. 1998. 49: 2273.
- [18] Nasibi, and KH. M- Kalantari. 2005. The Effectts of UV-A, UV-B and UV-C on protein and ascorbate content, lipid peroxidation and biosynthesis of screening compounds in *Brassica napus*. *Iranian Journal of Science & Technology, Transaction A*, Vol. 29, No. A1.
- [19] Ramawat, K. G.; Dass and Mathur. 2009. Herbal druges: ethno medicine to modern medicine. Springer, Berlin Heidelberg New York. 16.
- [20] Paula, C. and W. Virginia. 2004. Crosslinking of ribosomal proteins to RNA in maize ribosomes by UV-B and its effects on translation. *Plant physiology*, 136(2): 3319-3332.
- [21] Charles, M. L.; k. Tano; A. Asselin and Arul. 2008. Physiological Basis of UV-C induced resistance to *Botrytis cinerea* in tomato fruit V. Constitutive defence enzymes and inducible pathogenesisrelated proteins.
- [22] Burger, G. E. Edwards. 1996. Photosynthetic Efficiency and photodamage by UV and Visible Radiation, in Red versus Green Leaf Coleus Varieties, *plant cell physical*. vol.37, no. (3): 395-399.
- [23] Jordan, B. R.; W. S. Chow and J. M. Anderson. 1992. Changes in mRNA levels and polypeptide subunits of ribulose 1,5- bisphosphate carboxylase in response to supplemental UV-B radiation. *Plant, Cell and Environment*, 15: 91-8.
- [24] Hollosy F. 2002. Effects of ultraviolet radiation on plant cells. *Micron* 33: 179-197.
- [25] Zhang, J. E.; H. X. Henkow; L. Jorden and Strid, A. 1994. The effects of ultraviolet-B radiation on the CFoF₁-ATPASE. *Biochimica Biophysica Ada* 1185: 295- 302.