

Pharmacophore Modeling, Atom based 3D-QSAR and Docking Studies on ADAMTS-5 Inhibitors

Rathi Suganya P.

Assistant Professor, Department of Bioinformatics
SRM University
Kattankulathur, Chennai
E-mail: rathisuganya@ktr.srmuniv.ac.in

Sukesh Kalva

Department of Bioinformatics
SRM University
Kattankulathur, Chennai
E-mail: sukesh1411@gmail.com

Lilly M. Saleena

Assistant Professor, Department of Bioinformatics
SRM University
Kattankulathur, Chennai
E-mail: lmsaleena@ktr.srmuniv.ac.in

Abstract—ADAMTS-5 is an important aggrecanase that cleaves at key sites in the aggrecan core protein, in healthy and diseased cartilage. ADAMTS-5 deficient mice are protected from cartilage erosion in models of experimental arthritis. Therefore inhibition of ADAMTS-5 will be a potential cure for arthritis. In this study pharmacophore model was developed by downloading 50 ligands with IC50 value from BindingDB database. Pharmacophore Alignment and Scoring Engine (PHASE) software was used to develop ligand-based pharmacophore model for ADAMTS-5 using those 50 ligands. pIC50 ranged from 7.3149 to 5.018, of which pIC50 above 6.5 were considered as active and below 5.5 were considered as inactive. Three maximum hypotheses AAHRR, AARRR, AHRRR were generated. Pharmacophoric hypothesis AARRR.4144 had the best survival score of 3. 3D-QSAR was built for the best hypothesis with training set as 70% and atom based model was generated by keeping 1Å grid spacing and 6 as maximum number of PLS factors. Results show that AARRR.4144 has the best regression coefficient of 0.9832 and Pearson-R as 0.756. A docking study revealed the binding orientations of these inhibitors at active site amino acid residue His 373 of ADAMTS-5. The results of ligand-based pharmacophore hypothesis and atom based 3D-QSAR gave detailed structural insights as well as highlighted important binding features to design a novel therapeutically active compound against ADAMTS-5.

Keywords-3D-QSAR, pharmacophore, ADAMTS-5, Aggrecan, Docking.

I. INTRODUCTION

ADAMS (A Disintegrin And Metalloproteinase) is a peptidase protein which contains a unique integrin receptor-binding disintegrin domain, comes under the family of Metzincins. ADAMS are classified as Sheddases because they cut off or shed extracellular portions of transmembrane proteins. Two subfamilies are Snake venom metalloproteases (SVMPs) and ADAMTS (A disintegrin and

metalloproteinase with thrombospondin motifs) [2]. ADAMTS-4 and ADAMTS-5 are the major aggrecanase in human cartilage. ADAMTS-5 is synthesized in the rough endoplasmic reticulum, matures in the golgi compartment, constitutively expressed in human chondrocytes and synovial fibroblasts. ADAMTS-5 is an important aggrecanase that cleaves at key sites in the aggrecan core protein, in healthy and diseased cartilage. ADAMTS-5 deficient mice are protected from cartilage erosion in models of experimental arthritis [3]. Therefore inhibition of the ADAMTS-5 will be a potential cure for arthritis. Therapeutic effects of commercially available drugs last only for a short time due to their unfavourable pharmacokinetic profiles; therefore small target specific inhibitors could have enormous potential as new therapeutics. The overall fold of the catalytic domain resembles other metalloproteinases (MMP), but the shape of the substrate-binding site is unique. This unique binding site suggests that ADAMTS-5 recognizes different substrate motifs than MMP, ADAM and other ADAMTS enzymes. The unique binding site also increases the likely success of developing inhibitors that are specific for ADAMTS-5 [3].

Ligand-based drug designing approaches like pharmacophore mapping and quantitative structure-activity relationship (QSAR) are used in drug discovery. Database search studies for new hits and to identify important structural features for functional activity will help in identifying therapeutically stable drug without any side-effects [11].

In a rational drug design approach, identification of the pharmacophore is the most important step in achieving the stipulated goal. Pharmacophore Alignment and Scoring Engine (PHASE) software was used to develop ligand-based pharmacophore model for ADAMTS-5. PHASE uses conformational sampling and different scoring techniques to identify common pharmacophore hypothesis, each hypothesis is accompanied by a set of aligned conformations which are necessary for the ligand to bind to the receptor [9]

[10]. The developed model has the ability to find potential ADAMTS-5 inhibitors from 3D-virtual databases of drug-like molecules. The conformations of active compounds obtained from the alignment of pharmacophoric points are used to derive 3D-QSAR models. Further, the binding mode of the active molecule with the active site amino acid residues was performed by XP docking using Glide.

II. MATERIALS AND METHODS

A. Data set

For designing of novel potential ADAMTS-5 inhibitors, we downloaded 71 inhibitors available for ADAMTS-5 from the BindingDB database [3]. 55 compounds from the output had known IC₅₀ values. To avoid redundancy of information, the data set was further refined by removing compounds with similar biological activity and chemical structures by CANVAS (Schrodinger, LLC, New York, US) to identify diverse compounds, out of which we selected 50. The 50 compounds selected had IC₅₀ values of different range therefore the values (in moles/litre) were converted into negative logarithm of IC₅₀ (pIC₅₀). pIC₅₀ ranged from 7.3149 to 5.018, of which pIC₅₀ above 6.5 were considered as active and below 5.5 were considered as inactive and rests were moderately active.

B. Ligprep

These ligands were geometrically refined (cleaned) and conformers were generated with maximum number of conformers per structure as 1000 with force field OPLS-2005 with RMSD 1.0 Å°.

C. Hypothesis Generation

PHASE provides a standard set of six pharmacophore features, hydrogen bond acceptor (A), hydrogen bond donor (D), hydrophobic group (H), negatively ionizable (N), positively ionizable (P), and aromatic ring (R). Common pharmacophoric sites were selected from a set of variants and with the option Create Sites, number of acceptors were modified to 2, negatively ionizable to 0, others were kept default. This gave 6 different variant lists AAHRR, AAHRR, AARRR, AHRRR, AHRRR and HHRRR.

Hypothesis generation was done by Find option in find Pharmacophore model, which generated three maximum hypotheses with AAHRR, AARRR, AHRRR. For these hypothesis scores were calculated for both actives and inactives by score hypothesis using an overall maximum root mean square deviation (RMSD) value of 1.2 Å. The quality of alignment was measured by survival score.

D. 3D-QSAR

Phase provides the option of doing QSAR with the selected pharmacophore hypothesis. In the alignment option, align non-model ligands were chosen so that the ligands that are not part of the active set were also included. In Build QSAR option random training set was kept as 70% and atom based model was generated by keeping 1Å grid spacing and 6 as maximum number of PLS factors.

E. Molecular docking study

Docking was done using Glide (Schrodinger, LLC, New York, US). ADAMTS-5 crystal structure was downloaded from Protein Data Bank (PDB Id: 2RJQ) [16]. Protein was preprocessed, optimized and minimized with force field of OPLS2005 and RMSD of 0.30 Å using the protein preparation wizard. Grid was generated using the centroid of workspace ligand BAT. The docking was performed using Glide with enabling the “write XP descriptor information” option and keeping the rest default.

III. RESULTS AND DISCUSSION

ADAMTS-5 inhibitors can stop the expression of ADAMTS-5 thereby acting as a potential therapeutic drug for arthritis. In ligand based pharmacophore model we have developed a model which screened important pharmacophoric features necessary for these ligands to function as inhibitors. Training set consisted of 35 compounds, where 8 of them were active and 6 were inactive. Test set had 15 compounds. The pharmacophoric features selected for creating sites were hydrogen bond acceptor (A) and aromatic ring (R). Pharmacophore models containing three to five features were generated. The three and four featured pharmacophore hypotheses were rejected due to low value of survival score, as they were unable to define the complete binding space of the selected molecules. Five featured pharmacophore hypotheses was selected and subjected to stringent scoring function analysis.

102 different hypotheses were generated with AARRR, AHRRR and AAHRR; best 5 were shown in the table 1. Pharmacophoric hypothesis AARRR.4144 had the best survival score of 3.821. The pharmacophoric hypothesis of AARRR.4144 is shown in figure 1. The features represented in this hypothesis are two hydrogen acceptor and three aromatic rings. The distance and angles between the different sites are presented in the table 4 and 5 respectively. QSAR results also shows that the AARRR.4144 has the best regression coefficient of 0.9832, Pearson-R as 0.756. Result of atom-based 3D-QSAR with PLS 6 of AARRR.4144 hypothesis is shown in table 2.

The fitness score is checked for the pharmacophore model AARRR.4144. The best fitness score of 3 was with ligand number 10 (figure: 5). Best five fitness score compounds are shown in the table 3. Scatter plots for the predicted and experimental pIC₅₀ values for the ADAMTS-5 QSAR model applied to the training set and the test set are shown in figure 3 and 4 respectively.

A. 3D-QSAR Analysis

Inhibitory activity of the compound suggested by pharmacophore can be visualized by doing QSAR model. The results can be further used in designing novel ligands with the features derived from the pharmacophore model. The 3D-QSAR model was applied to the most active compound: 10 and the least active compound: 20, which are shown in the figure 6 and 7 respectively. These figures compare the most significant favourable (blue cubes) and

unfavorable (red cubes) regions for the activity of the compound.

B. Docking analysis

Extra precision glide docking (Glide XP) was performed for the best active compound 10 and ADAMTS-5(2RJQ). The docking results show interaction between compound 10 and ADAMTS-5 in the active site region with HIS373 with a G-Score of -9.14 kcal/mol (figure: 8). This complies with the 3D-QSAR model developed where the interaction is seen in the favourable region.

IV. CONCLUSION

In conclusion, developing a pharmacophore model will help in identifying therapeutically potential compounds without any side effects. Various pharmacophoric models were developed for ADAMTS-5 using 50 ligands downloaded from *BindingDB* database. Best hypothesis obtained was AARRR.4144 with two hydrogen bond acceptor and three aromatic rings. Compound 10 (sulfonfylamino-alkanecarboxylate, 38) had the best result for which a highly predictive atom based 3D-QSAR model was generated. Atom based 3D-QSAR and docking study helps in understanding the relationship between structure and activity. This gives us various options to design a novel and potent inhibitor for ADAMTS-5 protein.

ACKNOWLEDGMENT

Authors thank SRM University for their constant support and their funding for carrying out this study.

REFERENCES

- [1] Wolfsberg TG, Primakoff P, Myles DG, White JM, "ADAM, a novel family of membrane proteins containing A Disintegrin And Metalloprotease domain: multipotential functions in cell-cell and cell-matrix interactions.", *J Cell Biol.* 1995 Oct;131(2):275-8.
- [2] Seals DF, Courtneidge SA, "The ADAMs family of metalloproteases: multidomain proteins with multiple functions." *Genes Dev.* 2003 Jan 1;17(1):7-30.
- [3] Fosang AJ, Rogerson FM, East CJ, Stanton H, "ADAMTS-5: the story so far", *Eur Cell Mater.* 2008 Feb 5;15:11-26.
- [4] Cudic M, Burstein GD, Fields GB, Lauer-Fields J, "Analysis of flavonoid-based pharmacophores that inhibit aggrecanases (ADAMTS-4 and ADAMTS-5) and matrix metalloproteinases through the use of topologically constrained peptide substrates". *Chem Biol Drug Des.* 2009 Nov;74(5):473-82.
- [5] <http://www.bindingdb.org/bind/index.jsp>
- [6] Ujashkumar A. Shah, Hemantkumar S. Deokar, Shivajirao S. Kadam and Vithal M. Kulkarni, "Pharmacophore generation and atom-based 3D-QSAR of novel 2-(4-methylsulfonylphenyl)pyrimidines as COX-2 inhibitors.", 2010 Aug;14(3):559-568.
- [7] Tawari NR, Bag S, Degani MS, "Pharmacophore mapping of a series of pyrrolopyrimidines, indolopyrimidines and their congeners as multidrug-resistance-associated protein (MRP1) modulators.", *J Mol Model.* 2008 Oct;14(10):911-21.
- [8] Vipin Kumar*, Sunil Kumar and Poonam Rani, "Pharmacophore modeling and 3D-QSAR studies on flavonoids as α - glucosidase inhibitors.", *Der Pharma Chemica*, 2010, 2(4): 324-335.
- [9] Steven L. Dixon, Alexander M. Smondryev and Shashidhar N. Rao, "PHASE: A Novel Approach to Pharmacophore Modeling and 3D Database Searching.", *Chem Biol Drug Des* 2006; 67: 370–372.
- [10] Dixon SL, Smondryev AM, Knoll EH, Rao SN, Shaw DE, Friesner RA, "PHASE: a new engine for pharmacophore perception, 3D QSAR model development, and 3D database screening: 1. Methodology and preliminary results", *J Comput Aided Mol Des* (2006) 20:647–671.
- [11] Prashant Revan Murumkar, Vishal Prakash Zambre and Mange Ram Yadav, "Development of predictive pharmacophore model for in silico screening, and 3D QSAR CoMFA and CoMSIA studies for lead optimization, for designing of potent tumor necrosis factor alpha converting enzyme inhibitors", *Comput Aided Mol Des* (2010) 24:143–156.
- [12] Darren F. Seals and Sara A, "The ADAMs family of metalloproteases: multidomain proteins with multiple functions.", *Genes Dev* 2003 17: 7-30 .
- [13] Manas K. Majumdar, Roger Askew, et al, "Double-Knockout of ADAMTS-4 and ADAMTS-5 in Mice Results in Physiologically Normal Animals and Prevents the Progression of Osteoarthritis", November 2007, *ARTHRITIS & RHEUMATISM*, Vol. 56, No. 11, pp 3670–3674.
- [14] Matthew G. Bursavich,a, Adam M. Gilbert, et al. "Synthesis and evaluation of aryl thioxothiazolidinone inhibitors of ADAMTS-5 (Aggrecanase-2).", (2007), *Bioorganic & Medicinal Chemistry Letters* 17 1185–1188.
- [15] <http://www.pdb.org/pdb/home/home.do>

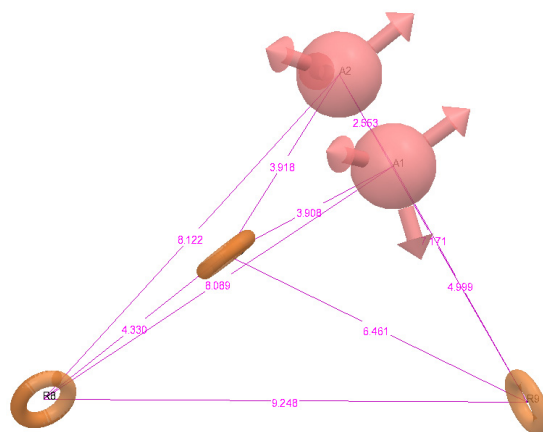


Figure 1. PHASE generated pharmacophore model AARRR.4144 illustrating hydrogen bond acceptor (A1, A2; pink), and aromatic ring (R8, R9, R10; orange) features with distances (in Å) between different sites.

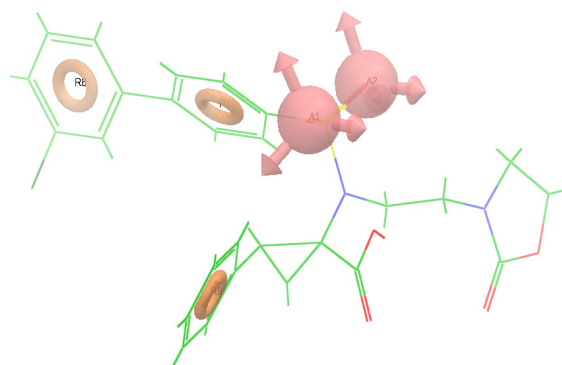


Figure 2. Best pharmacophore model AARRR.4144 aligned with molecule 10 illustrating hydrogen bond acceptor (A1, A2; pink), and aromatic ring (R8, R9, R10; orange)

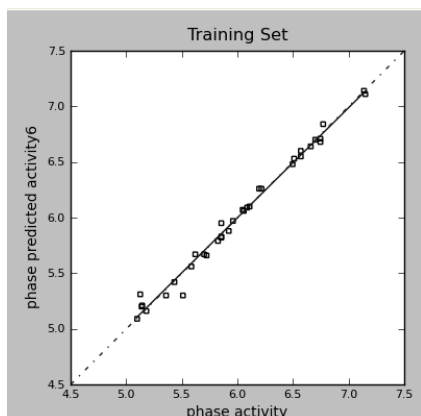


Figure 3. Scatter plots for the predicted and experimental pIC50 values for the ADAMTS-5 QSAR model applied to the training set.

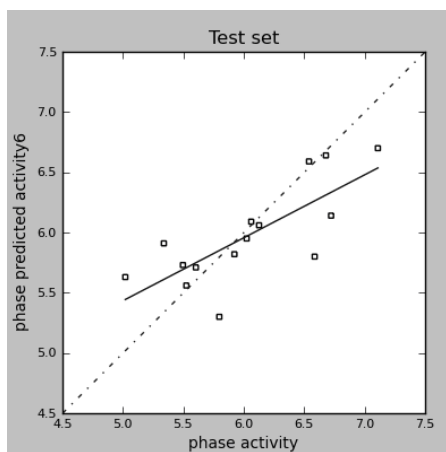


Figure 4. Scatter plots for the predicted and experimental pIC50 values for the ADAMTS-5 QSAR model applied to the test set.

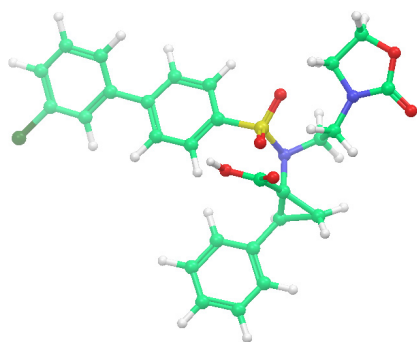


Figure 5. Compound 10: sulfonlamino-alkanecarboxylate, 38

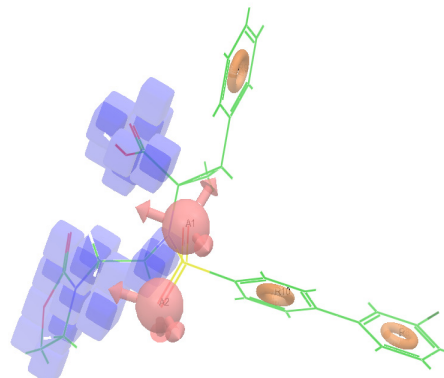


Figure 6. Atom based 3D QSAR model visualized in the context of most active compound 10. (Blue cubes indicate favorable regions while red cubes indicate unfavorable region for the activity)

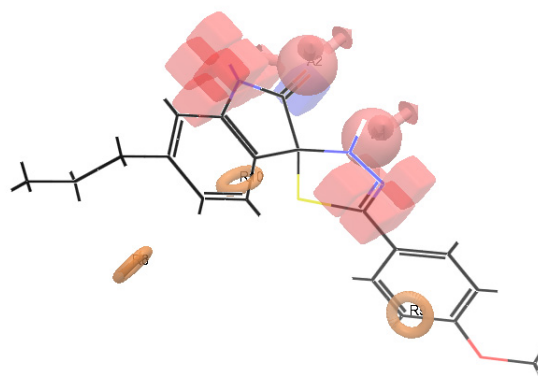


Figure 7. Atom based 3D QSAR model visualized in the context of least active compound 20.

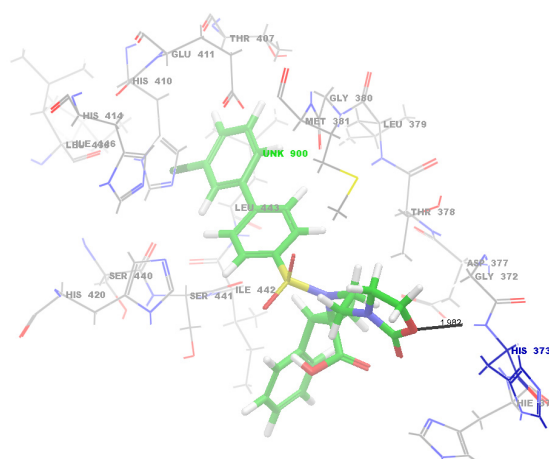


Figure 8. Docking of compound 10 in the active site of ADAMTS-5

TABLE I. BEST 5 HYPOTHESES GENERATED

S. No	ID	Survival score	Survival inactive score
1	AARRR.4144	3.821	2.296
2	AARRR.3936	3.798	1.987
3	AARRR.4159	3.774	2.282
4	AARRR.4283	3.749	1.953
5	AHRRR.2859	3.727	2.394

TABLE II. RESULTS OF ATOM-BASED 3D-QSAR WITH PLS 6 AARRR.4144 HYPOTHESIS.

ID	#	SD	R ²	RMSE	Q ²	Pearson-R
AARRR.4144	1	0.38	0.604	0.427	0.4455	0.6676
	2	0.2	0.887	0.4	0.5124	0.7286
	3	0.15	0.942	0.372	0.5789	0.7742
	4	0.13	0.96	0.369	0.5851	0.7783
	5	0.08	0.983	0.383	0.5532	0.756
	6	0.07	0.989	0.38	0.561	0.7593

SD = standard deviation of the regression, R² = correlation coefficient, Q² = for the predicted activities, RMSE = root-mean-square error, Pearson-R = correlation between the predicted and observed activity for the test set

TABLE III. THE BEST FIVE COMPOUNDS FOR AARRR.4144 HYPOTHESIS.

S. No	Compound #	QSAR Set	Experimental pIC50	Predicted pIC50	Fitness
1	10	training	6.658	6.82	3
2	5	training	6.745	6.79	2.97
3	13	training	6.569	6.75	2.97
4	9	test	6.678	6.78	2.95
5	12	training	6.569	6.76	2.95

TABLE IV. THE DISTANCE BETWEEN THE DIFFERENT SITES OF AARRR.4144 HYPOTHESIS

Site1	Site2	Distance
A1	A2	2.553
A1	R8	8.089
A1	R9	4.999
A1	R10	3.908
A2	R8	8.122
A2	R9	7.171
A2	R10	3.918
R8	R9	9.248
R8	R10	4.33
R9	R10	6.461

TABLE V. THE ANGLES BETWEEN THE DIFFERENT SITES OF AARRR.4144 HYPOTHESIS

Site1	Site2	Site3	Angle
A2	A1	R8	81.7
A2	A1	R9	141.3
A2	A1	R10	71.2

R8	A1	R9	86.5
R8	A1	R10	11.5
R9	A1	R10	92.2
A1	A2	R8	80.2
A1	A2	R9	25.8
A1	A2	R10	70.7
R8	A2	R9	74.1
R8	A2	R10	10.6
R9	A2	R10	63.6
A1	R8	A2	18.1
A1	R8	R9	32.7
A1	R8	R10	10.4
A2	R8	R9	48.2
A2	R8	R10	9.6
R9	R8	R10	38.7
A1	R9	A2	12.9
A1	R9	R8	60.8
A1	R9	R10	37.2
A2	R9	R8	57.6
A2	R9	R10	32.9
R8	R9	R10	24.8
A1	R10	A2	38.1
A1	R10	R8	158.2
A1	R10	R9	50.6
A2	R10	R8	159.9
A2	R10	R9	83.6
R8	R10	R9	116.6