

Diacetylmonoxime Reactivation of Acetylcholinesterase and Butyrylcholinesterase Inhibited by Dichlorvos in Central and Peripheral Nervous System of Rat

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Abstract. The *in vivo* AChE and BuChE inhibition in different tissues of the male albino rats by an organophosphorus compound dichlorvos (DDVP; 47 mg/kg) was examined. The AChE percent inhibition in the brain regions was as follows: striatum>hippocampus>medulla>pons>cortex>thalamus>cerebellum>spinal (65.6>55.3>52.4>45.6>41.6>35>22.5>16.6). In the other target organs the AChE inhibition was in the order of whole brain>liver>kidney>testis>serum (40> 39.9>38>37>30>29.2). *In vivo* inhibition of serum BuChE was more severe than inhibition of its AChE. In the kidney AChE activity was more inhibited in comparison to its BuChE activity. The *in vivo* effect of pretreatment of DAM (100 mg/kg) on DDVP-induced AChE inhibition was also studied. DAM provided significant protection ($p < 0.01$) against DDVP inhibited AChE in almost all tissues. DAM protected 90 % ($P < 0.01$) against DDVP inhibition in pons and spinal, about 75 % in hippocampus, medulla and cortex. DAM did not show significant protection against serum AChE inhibition but provided significant ($p < 0.01$) protection against serum BuChE inhibition.

Keywords: AChE, BuChE, Dichlorvos, DDVP, DAM, Inhibition, Reactivation, Rat, Brain.

1. Introduction

Despite the use of new selective insecticides organophosphorus (OPs) remains an important class of pesticides. Dichlorvos, an organophosphorus compound (DDVP: 2,2-dichlorovinyl dimethyl phosphate) is a contact and stomach-acting insecticide with fumigant and penetrant action. It is widely used as crop protection against sucking and chewing insects. Non-target species including mammals are frequently injured by OPs (1). Neurotoxicity from OP exposure is generally related to inhibition of acetylcholinesterase (AChE; EC 3.1.1.7), (2). Which subsequently result in the accumulation of Acetylcholine (ACh) at neuroeffector sites (3). Inhibition of brain AChE is generally regarded as a useful marker and most sensitive measure of organophosphorus toxicity (4). Regarding the neurochemical mechanism of OPs effects, a study on discrete brain regions (cortex, cerebellum, striatum, hippocampus, stem, thalamus, pons, and optic chiasma) serum, kidney, and testis showed a considerable quantitative difference of inhibition of AChE (5). All OPs elicit their primary effects by phosphorylating or phosphonylating the serine hydroxyl at the active site of the enzyme AChE, causing accumulation of excess ACh at various cholinergic sites causing toxic manifestations (6). The inhibited enzyme can be reactivated by certain drugs (cholinesterase reactivators) such as hydroxylamine, hydroxamic acids and oximes. (7). Among the oximes pralidoxime (PAM), obidoxime and trimedoxime (TMB-4) are well known cholinesterase reactivators (8,9). Recently H-oxime HI6 and HLÖ7 are proved to be most effective oxime against toxic nerve agents (10,11). Only a few studies conducted recently have offered some information regarding the neurochemical mechanism of reactivation of the effect of OPs by oximes in brain regions and other tissues. The present study was undertaken to evaluate the ability of Diacetylmonoxime (DAM) to reactivate Rat AChE inhibited by DDVP in discrete regions of the brain, Serum, liver, kidney, testis and whole brain. Butyrylcholinesterase (BuChE; EC 3.1.1.8)

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is present in the central nervous system in appreciable amount but appears to be associated mainly with supportive elements such as glia or vascular tissue, without direct implications in neurotransmission (12). Effect of DAM on DDVP-induced BuChE inhibition in serum and kidney was also studied.

2. Materials and Methods

2.1. Chemicals

Acetylthiocholine iodide (ATCI), Butyrylthiocholine iodide (BuTCI), 5,5'-Dithio-bis 2-nitrobenzoic acid (DTNB) and bovine serum albumin were purchased from Sigma chemicals CO. (St. Louis M.O.). Technical grade dichlorvos, (DDVP; 2,2-dichlorovinyl dimethyl phosphate) and Diacetylmonoxime (DAM) were obtained from All India Medical Corporation (Mumbai India). All other chemicals used were of analytical grade.

2.1.1 Animals & Treatment

Male albino Rats (CFT-Wistar strain) weighing between 225-250 g were used for these investigations. Rats were maintained in individual plastic cages in a temperature-controlled room under standard laboratory conditions with free access of water and commercial feed ad lib. Rats were divided into four groups of five each. Experimental rats were fasted overnight. Each group receiving different treatment(s): Group I: Control animals; oil (1 ml/kg). Group II: single dose DDVP (47mg/kg; 1/3 LD₅₀), Group III: DAM (100mg/kg) as solution in isotonic saline i.m. Injection volume was 1ml/kg. Group IV: DAM (100 mg/kg) i.m. pre-treatment, 30 min. before, DDVP (47mg/kg) administration. Rats in groups III and IV received daily DAM (100mg/kg) for three days. Stock solution of DDVP (16mg/ml) was prepared in 100% pure peanut oil. Stock solution of DAM (100mg/ml) was prepared in normal saline. Stock solutions were made freshly just before the use.

2.1.1.1 Sample preparation & Enzyme assay

Rats (group of five) were anaesthetized by Diethyl ether 16 hr. after administration of DDVP and their abdominal cavities surgically opened. For serum sample blood was collected by cardiac puncture. Blood was allowed to clot for 5 min. followed by centrifugation at 1000 x g at 4°C for 10 minute. Brains were then quickly removed, washed in ice-cold saline and blotted dry. Each brain was placed on ice bag and different regions viz. cortex, striatum, cerebellum, pons, thalamus, hippocampus, medulla and spinal cord were isolated according to Zeman and Innes (13). Liver, kidney and testis were also taken out immediately. A 10 % (w/v) tissue homogenate was prepared in 0.1 M phosphate buffer (pH 7.4) using potter-Elvehjem type homogenizer followed by centrifugation at 2000 x g at 4°C for 10 min. in a sorwell high speed refrigerated centrifuge. All homogenates were kept at -70°C immediately after preparation and analyzed on the same day for the activity of AChE and BuChE. AChE activity was determined spectrophotometrically by measuring the rate of hydrolysis of the substrate, ATCI/BuTCI according to the method of Ellman *et al.*, (14). Aliquots of the homogenates were diluted with 2.6 ml 0.1 M phosphate buffer (pH 7.4) to a total volume of 3 ml, which contained 100 mM DTNB and 75 mM ATCI or BuTCI. The enzyme activity was calculated by extinction co-efficient ($E_{412} = 1.36 \times 10^4 \text{ M}^{-1} \text{ cm}^{-1}$) for the yellow anion 5-thio-2 nitrobenzoic acid. The AChE and BuChE activity was expressed as nmole of ATCI / BuTCI hydrolyzed per min per milligram of protein and for serum ATCI/BuTCI hydrolyzed/ml serum/min. All spectrophotometric measurements were done in duplicate. Proteins were determined according to Lowry *et al.* (15) using bovine serum albumin as standard.

2.1.1.1.1. Statistical analysis

LD₅₀ values were calculated by probit regression analysis of death occurring within 24hr. after administration of DDVP at five different doses with six Rats per dose (16). Mean values and standard deviation were calculated for each test group on the basis of value obtained for individual tissue from Rats. Data were analyzed by one-way ANOVA to determine treatment effects. In all cases $p > 0.05$ or $p > 0.01$ was considered statistically significant.

3. Results

3.1. Control Enzyme Activity

Baseline enzyme level for AChE is shown in Table –1 and –2. Result shows differential AChE activity in discrete regions of the brain. Among the brain regions the highest AChE activity in striatum was 3.673 ± 1.2 nmole/min/mg protein and the lowest in the hippocampus and thalamus was 1.857 ± 6.2 and 1.880 ± 0.9 nmole/min/mg proteins respectively. The discrete regions of control rat brain showed differences in the AChE activities in the order of striatum > medulla > pons > cortex > cerebellum > hippocampus > thalamus > spinal. In the other organs AChE level was in the order of serum > brain > kidney = testis > liver. The serum BuChE activity (12.92 ± 4.9 nmole/min) was almost four times more than its AChE activity (3.792 ± 4.3 nmole/min). Similarly in the kidney also the level of BuChE was much higher i.e. 4.741 ± 7.2 nmole/min/mg protein in comparison to its AChE activity which was 1.591 ± 2.7 nmole/min/mg protein.

3.1.1 *In vivo* inhibition of AChE and BuChE by DDVP

Single dose of DDVP (47mg/kg) elicit differential toxicity in the brain region. The *in vivo* inhibition of AChE in the discrete regions of rat brain ranged from 65.5% (in striatum; $p < 0.01$) to 22.5% (in cerebellum; $p < 0.05$) (Fig- 1). DDVP elicit significant percent inhibition in AChE activity of the brain regions in the order of striatum > hippocampus > medulla > pons > cortex > thalamus > cerebellum > spinal ($65.6 > 55.3 > 52.4 > 45.6 > 41.6 > 35 > 22.5 > 16.6$). AChE inhibition was also significant ($p < 0.05$) in other organs due to DDVP and was in the order of liver > kidney > whole brain > testis > serum ($39.9 > 38 > 37 > 30 > 29.2$; Fig-2). In the serum DDVP-induced BuChE inhibition (38.9 %) was more pronounced than was its AChE inhibition (29.2 %). However in the kidney AChE activity was more inhibited than was its BuChE activity.

3.1.1.1 *In vivo* effect of DAM on AChE

Administration of DAM alone (100 mg/kg) to rats for three days did not produce any significant inhibition of AChE in the brain regions and the other target organs hence efficacy of DAM on DDVP-induced AChE inhibition was calculated from control values (Fig-3 & 4).

3.1.1.1.1 Effect of pretreatment of DAM against DDVP-induced AChE and BuChE inhibition

The Data in Table-1 indicate that the pretreatment of DAM (100 mg/kg) for three days prior to the single dose of DDVP (47 mg/kg) to the male rats provided significant protection ($p < 0.01$) against DDVP inhibited AChE in almost all regions of the brain. Results from the study revealed that DAM offered maximum protection with more than 90% ($p < 0.01$) reactivation in pons and spinal. Although it provided more than 75% protection against DDVP inhibited AChE in the hippocampus, medulla and cortex. However, DAM produced 50% protection in the cerebellum and thalamus and 40% protection in the striatum against DDVP inhibited AChE (Fig- 3). DAM did not show any significant protection against AChE inhibition in the serum but provided highly significant ($p < 0.01$) protection against inhibition of serum BuChE (Table-2). Highest protective action of DAM was found significantly ($p < 0.01$) in reactivating AChE inhibition in the testis, which was reactivated to normal level of AChE. In case of liver AChE inhibition DAM also showed considerable protection but unable to produce any protection in the kidney (Table-2 & Fig-4).

4. Discussion

The present investigations demonstrate protective efficacy of Diacetylenoxime against AChE inhibition produced by DDVP in discrete regions of the brain in addition to whole brain, liver, kidney, testis and serum. The regional difference in the level of AChE of brain areas of untreated rat was significant ($p < 0.05$) in the present investigation. Highest AChE activity was found in the striatum and lowest in the hippocampus and thalamus. However, lowest level of AChE was reported in the cerebellum (17). Striatum is the area rich in cholinergic neurons and AChE activity was higher in this area (18). The differential AChE activity in the brain regions of untreated rats was, in the present study, in confirmation (5) except we found lower AChE activity in the thalamus than in the cortex and cerebellum. AChE activity was comparatively found to be lower in serum, kidney and testis. It has been reported that in Kidney, liver and serum AChE activity is low (19). BuChE activity was found to be the predominant than AChE activity in serum and kidney of untreated rats. This may account for higher specificity to the substrate BuTC compared to ATC (20). BuChE in serum samples of mammals including rat showed more preference to BuTC compared to ATC (21). Upon inhibition of AChE by organophosphorus in the nervous system, the neurotransmitter acetylcholine accumulates in synapses, causing excessive stimulation of cholinergic receptors on

postsynaptic cells, leading to cholinergic toxicity. It is known that feedback inhibition of acetylcholine release can occur through activation of muscarine acetylcholine receptors located on presynaptic terminals (22, 23). Presynaptic muscarinic receptors diminish further Acetylcholine release and thereby may reduce the excessive stimulation of postsynaptic cholinergic receptors following AChE inhibition (24). Present study reveals that DDVP produce maximum toxicity in brain regions and whole brain. AChE inhibition was 65% in striatum, 55% in hippocampus, 52% in medulla, 45% in pons, 41% in cortex, 35% in thalamus, 22% in cerebellum and 16% in spinal. These findings were similar since soman as previously reported to produce maximum inhibition of AChE in the striatum of rat (25). Regional differences in AChE inhibition by organophosphorus depends on their adaptive responses to pesticide exposure could therefore influence sensitivity to some anticholinesterase (26). It has been stated that brain is the first organ affected by organophosphorus poisoning (27) and an early inhibition of AChE takes place in this organ (28). The spinal cord is often neglected portion of CNS. Yet we have shown that DDVP affects AChE activity to the same extent here as in the major brain regions (29). Excellent correlation between brain regional and spinal cord AChE activity was reported (30, 31). AChE inhibition in either in nervous tissues or muscles has fairly been accepted as a toxic effect because activity in these target tissues is involved in neurotransmission (32). But AChE inhibition by OPs on other tissues or organs (such as liver, kidney and testis) different from them has seldom been studied. Results of this investigation of DDVP poisoning in rat AChE activity was inhibited significantly ($p < 0.05$) i.e. 40% in liver, 38% in kidney and 30% in testis. The differential inhibition of AChE activities in these tissues may be due to presence of isoenzyme with different affinity for the substrate and the inhibitor. Further pesticide itself may be present in different amount in different tissues producing differential inhibition or inhibitor may be metabolized at different rates (33). Regarding the differential inhibition of serum AChE and BuChE in DDVP treated rats BuChE appeared to be slightly more inhibited than was AChE. The inhibition of BuChE can serve as protective mechanism against DDVP toxicity by reducing the amount available for AChE inhibition (18).

The result of the present investigation demonstrates a role for the direct action of DAM in reactivating AChE inhibition after exposure of rats to single dose of DDVP. Pretreatment of DAM provided protection against DDVP-induced AChE inhibition in all of the brain areas examined; however each area was not affected to the same degree. DAM exhibited excellent protection against the toxic effect of DDVP (demonstrating the AChE reactivation) produced nearly equivalent levels of protection in hippocampus, pons and medulla. The amount of reactivation of phosphorylated brain enzyme that can be achieved by TMB-4 or toxogonin differ in different areas of brain, due to regional differences in the amount of functional (i.e. more accessible, extra cellular) enzyme and non-functional (i.e. less accessible, intracellular) enzyme (34). Kuca and co-workers (35) reported that there were significant differences in reactivation potency of all tested oximes. The oxime TO205 seems to be the most efficacious followed by TO046, HI-6, HS-6, K027, obidoxime, MMC and 2-PAM. In addition, the findings of their study revealed that the reactivation potency of the tested reactivators depends on many factors such as the number of pyridinium rings, the number of oxime groups and their position, as well as the length and the shape of Linkage Bridge between two pyridinium rings.

Variation in reactivation of Tabun induced AChE inhibition has been reported by three oximes HI-6, obidoxime, and K048 (36). Patocka (11) showed DAM was about 10 times more effective against sarine poisoning in rat than in mouse, guinea-pig, rabbit and monkey. It has been reported that MINA and DAM were effective in reducing AChE inhibition by organophosphorus pesticides (37). It is known that atropine sulphate is essential part of the treatment of OP poisoning (38). It is considered that HI-6 is more effective than 2-PAM in the therapy of OP toxicity, especially for agents that undergo rapid aging (39). The difference may be attributed, at least in part to higher reactivating potency of HI-6 (40, 41).

In conclusion, prophylactic administration of DAM potentially protected AChE activity in brain regions in addition to serum and other tissues from inhibition by DDVP. One possible explanation for this might be slowing of aging of DDVP-AChE complex. Alternately, DAM might block the binding of DDVP to the esteric site of AChE (12). DAM offers possibility of providing prophylactic benefit against DDVP toxicity. Thus DAM may prove useful therapy of DDVP intoxication.

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6. References

- [1] D. M. Maxwell, K.M. Brecht, F.C.T. Chang, I. Koplovitz, T.M. Shih and R.E. Sweeney. Toxicodynamic modeling of highly toxic organophosphorus compounds. *J. Mol. Neurosci.*, 2006, **30**, 129-132.
- [2] W.N. Aldridge. Mechanism and concept in toxicology. Tayler & Francis (Eds.), London. pp. 86-90. (1996).
- [3] J. Bajgar. Organophosphates/nerve agents poisoning; mechanism of action, diagnosis, prophylaxis and treatment. *Adv. Clin. Chem.* 2004, **38**, 151-216.
- [4] V. Racakova, .D. Jun, V. Opletalova, & K. Kuka. Reactivation of acetylcholinesterase inhibited by the pesticide chlorpyrifos. *J. Appl. Biomed.* 2006, **4**, 147-151.
- [5] P. Santosh kumar, S. Karanth & T. Shivanandappa. Neurotoxicity and pattern of acetylcholinesterase inhibition in the brain regions of rat by bromophos and ethylbromophos. *Fundamental and Applied toxicology*, 1996, **32**, 23-30.
- [6] D.F. Heath. Organophosphorus poisons. Anticholinesterase and related compounds. *Pergamon London, 1961* 17-28.
- [7] V. Racakova, M. Hrabnova, D. Jun & K. Kuca. Substituted monoquarternary oximes as reactivators of cyclosarin and Chlorpyrifos- inhibited acetylcholinesterase. *Arh. Hig Rada Toksikol*, 2006, **57**, 387-390.
- [8] D.M. Maxwell, I. Koplovitz, F. Worek and R.E. Sweeney. A structure-activity analysis of the variation in oxime efficacy against nerve agents. *Toxicol. Appl. Pharmacol.*, 2008, **231**, 157-164.
- [9] J. Caval, K. Kuca & J. Kassa. Specification of structure of oximes able to reactivate tabun-inhibited acetylcholinesterase. *Basic Clin. Pharmacol. Toxicol.*, 2004, **95**, 81-86.
- [10] P. Eyer, I. Hegedorn, R. Climmek, P. Lipstreu, H. Oldiges, I. Steidl, I. Szinicz & T.Worek. HIÖ7 Dimethane a potent bispyridinium dioxime against anticholinesterase. *Aech. Toxicol.* 1992, **66**, 603-621.
- [11] J. Patocka, J.Cabal, K. Kuca, & D. Jun. Oxime reactivation of acetylcholinesterase inhibited by toxic phosphorus esters: in vitro kinetics & thermodynamics. *J. Appl. Biomed*, 2005, **3**, 91-99.
- [12] A. Silver. Anticholinesterases. In *The Biology of Cholinesterases*. A. Euburger and E. I. Tatum..Eds. North-Holland. Amsterdam, 1974, 464-471.
- [13] W. Zeman, J.R.M. Innes. Craigie's Neuroanatomy of the Rat. 1963, 21-25, Academic Press. New York.
- [14] G.L. Ellman, K.D. Courtney, V. Anders. & R.M. Featherston. A new and rapid colorimetric determination of acetylcholinesterase activity. *Biochem. Pharmacol.* **7**, 1961. 88-95.
- [15] O.H. Lowry, N.J. Rosenbrough, A.L. Farr, J. Randall. Protein measurement with the folin-phenol reagent. *J. Biol. Chem.* **193**, **1951**, 265-275.
- [16] D.J. Finny. Probit Analysis, 1977, 50-80, Cambridge Univ. Press Cambridge.
- [17] E.L. Bennet, M.C. Diamond, H. Morimoto. & M. Herbert. Acetylcholinesterase activity and weight measures in fifteen brain areas from six lines of rats. *J. Neurochem.* **13**, 1966, 563-581.
- [18] H. Michalek, & W.B. Stavinoha. Effect of chlorpromazine pretreatment on the inhibition of total cholinesterase and butyrylcholinesterase in brain of rats poisoned by physostigmine or dichlorvos. *Toxicology.* **9**, 1978, 205-219.
- [19] K.B. Augustinsson., Cholinesterases. A study in comparative enzymology. *Acta Physiol Scand.* 1948, **52**, 1-182.
- [20] C.H. Sawyer & J.W. Everett. Cholinesterase in rat tissue and site of serum non-specific cholinesterase production. *Am. J. Physiol.* 1947, **148**, 675-683.
- [21] K. Kuca, J.Patocka & Cabalj. Reactivation of organophosphate inhibited acetylcholinesterase activity by α, ω -bis-(4- hydroxyiminomethylpyridinium) alkanes in vitro *J. Appl. Biomed.* 2003, **1**, 307-211.
- [22] C. Allgaier, B.K. Choi & G. Hertting. Muscarine receptors regulating electrically evoked release in acetylcholinesterase in hippocampus are linked to pertussis toxin- sensitive G protein but not to adenylate cyclase. *J.*

Neurochem. 1993, **61**, 1043-1049.

- [23] T.W. Vikroy & E.D. Cadman. Dissociation between muscarinic receptor-mediated inhibition of adenylate cyclase and autoreceptor inhibition of [3H] acetylcholine release in rat hippocampus. *J. Pharmacol. Exp. Ther.* 1989, **251**, 1039-1044.
- [24] C. Pope. The influence of age on pesticide toxicity. Hand book of pesticide toxicology. Academic press. **1**, 2001, 873-885.
- [25] D.K. Lim, A.B. Porter, B. Hoskins & K. Ho. Changes in ACh levels in rat brain during sub acute administration of Di-isopropylefluorophosphate. *Toxicol. Appl. Pharmacol.* 1987, **90**, 477-489.
- [26] K.V. swamy, R. Ravikumar & M. Mohan. Changes in cholinesterase system in different brain areas during the development of behavioural tolerance to monocrotophos toxicity in male albino rats. *Biochemistry International* 1992, **27 (4)**, 661-669.
- [27] F. Worek, T. Kirchner, M. Backer, L. Szinicz. Reactivation by various oximes of human erythrocyte acetylcholinesterase inhibited by different organophosphorus compounds. *Arch. Toxicol.* 1996, **70**, 497-503.
- [28] A.C. Rosenfeld & L.G. Sultatos. Concentration-Development Kinetics of acetylcholinesterase inhibition by the organophosphate Paraoxon. *Toxicological sciences.* 2006, **90 (2)**, 460-469.
- [29] V.R. Jimmerson, T.M. Shih, M. Pennella, T. Koviak, O. Smith, F. Cowan & R.B. Mailman. Spinal cord cholinesterase activity predicts brain regional cholinesterase activity in soman (GD) poisoned rats. *Toxicologist.* 1986, **6**, 194-201.
- [30] V.R. Jimmerson, T.M. Shih & R.B. Mailman. Variability in soman toxicity in the rat correlation with biochemical and behavioural measures. *Toxicology.* 1989, **57**, 241-254.
- [31] T.M. Shih, W. Jacob, J. C. Skovira, O'Donnell & J. H. McDonough. Central Acetylcholinesterase Reactivation by oximes improves survival and terminates seizures following nerve agent intoxication advanced studies in biology, **1(3)**, 2009, 155 – 196.
- [32] S. Padilla, J. Buzzard, & V.C. Moser, V.C., Comparison of the role of Esterase in the age related sensitivity to chlorpyrifos and Methidamidophos, *Neurotoxicology* 2000, 49-56.
- [33] T. Shivanandappa, P. Joseph & M.K. Krishnakumari. Response of blood and brain cholinesterase to dermal exposure of bromophos in the rat. *Toxicology.* **4**, 1998, 199-208.
- [34] F. Hobbiger & V. Vojvodic. The reactivation by pyridinium aldoxime of phosphorylated acetylcholinesterase in the central nervous system. *Biochem. Pharmacol.* 1967, **16**, 455-459.
- [35] K. Kuca, L. Musilova, J. Palecek, V. Cirkva, M. Paar, K. Musilek, M. Hrabanova, M. Pohanka, J. Z. Karasova and D. Jun. Novel Bisquaternary Oximes—Reactivation of Acetylcholinesterase and Butyrylcholinesterase inhibited by Paraoxon. *Molecules*, **14**, 2009, 4915-4921.
- [36] J. Bajgar, J.K. Kassa, A. Paseka, D. Slizova, O. Krs K, Kuca, D. Jun, J. Fusek & L. Capek. A comparison of tabun-inhibited rat brain acetylcholinesterase reactivation by three oximes (HI-6, obidoxime, and K048) in vivo detected by biochemical and histochemical techniques. *J Enzyme Inhib Med Chem.* 2010, **25 (6)**: 790-797.
- [37] D.R. Davis. & A.L. Green. The chemotherapy of poisoning by organophosphate anticholinesterases. *Brit. J. Indust. Med.* **16**, 1959, 128-132.
- [38] K. Musilek, K. Kuca, D. Jun & M. Dolezal. Progress in synthesis of new acetylcholinesterase reactivators during the period 1990-2004. *Curr. Org. Chem.* **11**, 2007, 229-238.
- [39] M. Alder, D.M. Maxwell, R.E. Sweeny & S.S. Deshpande. Evaluation of the direct action of HI-6 in reversing soman-induced titanic fade. In. *Enzymes of the Cholinesterase Family.* M. Danial. Quinn et al. Eds. Plenum Press. New York. 1995, 321-329.
- [40] M. Calic, A. Lucic-Vrdoljak, B. Radic, D. Jelic, D. Jun, K. Kuka & Z. Kovarik. In vitro and in vivo evaluation of pyridinium oximes, mode of interaction with acetylcholinesterase, effect on tabun-and soman-poisoned mice and their cytotoxicity. *Toxicology*, 2006, 219 85-96, 2006.
- [41] S. Karina, Matos, T. Daiana. Mancini, F.F. Elaine, da Cunha, K. Kuča, C.C. Tanos, C. França & T. Ramalho. Molecular aspects of the reactivation process of acetylcholinesterase inhibited by cyclosarin. *J. Braz. Chem.*

Table 1: Effect of pretreatment of Diacetylmonoxime (DAM) on DDVP- induced acetylcholinesterase inhibition in the discrete regions of Rat brain.

AChE activity n mole/ATCI hydrolyzed/min/mg protein				
Brain regions	Control	DDVP	DAM	DAM+DDVP
Striatum	3.673±1.2 (100)	1.261±3.4** (34.4)	3.432±6.4 (93.4)	2.213±1.7* (60.3)
Hippocampus	1.857±6.2 (100)	0.83±9.4** (44.6)	1.723±2.6 (93.2)	1.692±7.1** (91.1)
Medulla	3.538±0.37 (100)	1.685±0.98* (47.6)	3.39±1.3 (95.9)	3.22 ±4.2** (91.1)
Pons	2.667 ±1.1 (100)	1.453±12.4** (54.5)	2.592±2.8 (97.1)	2.745±1.4** (102.9)
Cortex	2.212±1.9 (100)	1.3±4.3* (58.4)	2.114±8.1 (95.5)	1.966±3.4** (88.8)
Thalamus	1.88±0.9 (100)	1.225±3.6* (65)	1.886±7.17 (99.1)	1.528±6.9* (81.2)
Cerebellum	2.159±0.15 (100)	1.675±8.1* (77.5)	2.071±9.2 (95.9)	1.973±2.7** (89.8)
Spinal	1.774 ± 1.8 (100)	1.479±4.4 (83.4)	1.716±0.6 (96.7)	1.667±13.1 (98.4)

Note: Data represent mean± S.E. (n = 5 groups). Asterisk(s) indicates a significant difference * P<0.05 and ** < 0.01 between treated and control, the numerals in parentheses denote the % change from control.

Table 2: Effect of pretreatment of Diacetylmonoxime (DAM) on DDVP – induced AChE and BuChE inhibition in the various tissues of Rat.

AChE activity nmole/ATCI hydrolyzed/min/mg protein				
Tissues	Control	DDVP (47mg/kg)	DAM (100mg/kg)	DAM+DDVP
Serum ^a	3.792±4.3 (100)	2.686±11.2* (70.8)	3.697±17.6 (97.4)	2.945±9.6* (77.6)
Whole brain	2.164±2.9 (100)	1.36±3.1** (60)	2.084±2.4 (96.3)	1.858±1.5 (85.8)
Testis	1.695±19 (100)	1.179±8.2* (69.5)	1.973±6.5 (116.4)	1.929±9.2** (113.8)
Kidney	1.591±2.3 (100)	0.979±6.2± (62)	1.546±12 (96.9)	1.121±14.5 (70)
Liver	1.17±15 (100)	0.704±9.8* (60)	1.191±3.3 (101.7)	0.989±17 (84.5)
Serum (BuChE) ^a	12.92±6.9 (100)	7.9±14* (61.1)	12.72±17.6 (98.4)	12.58±21.3** (97.3)
Kidney (BuChE) ^b	4.7±12 (100)	3±4.9* (73.4)	4.68±8.3 (97.8)	3.588±6.4 (76.6)

Note: * Data represents mean ± S.E. (n = 5 groups). Asterisk(s) indicates a significant difference * P<0.05 and ** < 0.01 between treated and control, the numerals in parentheses denote the % change from control. (a) Activity was expressed as nmole substrate hydrolyzed ml serum⁻¹ min⁻¹ (b) Butyrylcholinesterase activity expressed as nmole BuTCl hydrolyzed/min/mg protein.

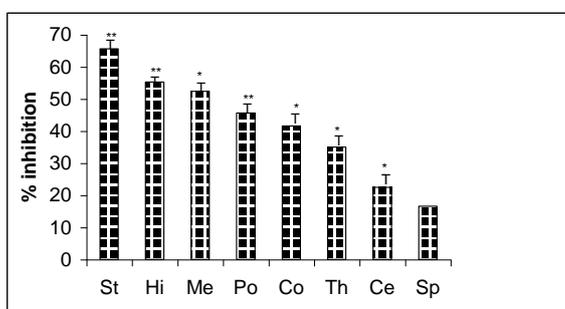


Fig. 1: DDVP-induced AChE inhibition in brain regions of Rat. Results are expressed as percentage of inhibition (as compared with untreated rats) ± S.E.(St. striatum; Hi. hippocampus; Me. medulla; Po. Pons; Co. cortex; Th. Thalamus; Ce. Cerebellum; Sp. Spinal.). *Significance difference between treated and control groups * P<0.05 and ** P<0.01.

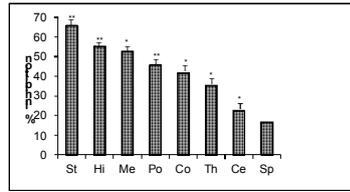


Fig. 2: DDVP-induced AChE inhibition in various tissues of Rat. Results are expressed as percentage of inhibition (as compared with untreated rats) \pm S.E. (Wb. Whole brain; Liv. Liver; Kid. Kidney; Tes. testis; ser. Serum)
 *Significance difference between treated and control groups * $P < 0.05$ and ** $P < 0.01$.

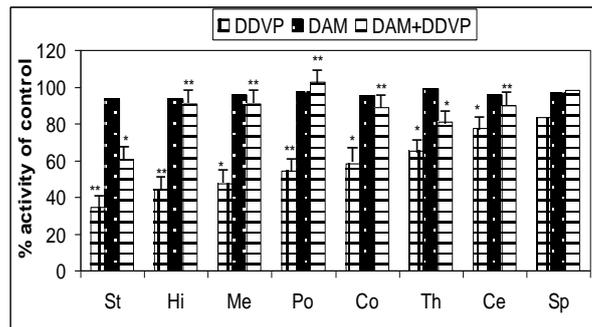


Fig. 3: Effect of pretreatment of DAM (100mg/kg; im. For three days prior to administration of 47mg/kg DDVP) and DDVP on AChE activity (*in Vivo*) in discrete regions of rat brain.(St. striatum; Hi. hippocampus; Me. medulla; Po. Pons; Co. cortex; Th. Thalamus; Ce. Cerebellum; Sp. Spinal.). *Significance difference between treated and control groups * $P < 0.05$ and ** $P < 0.01$.

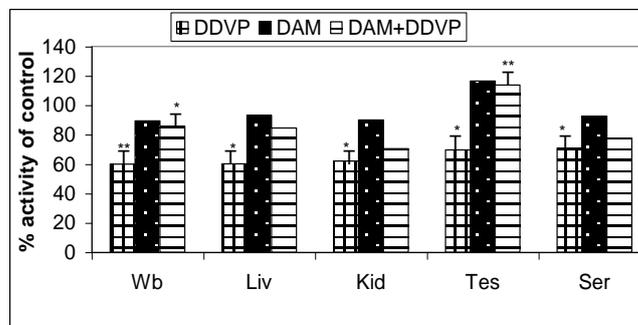


Fig. 4: Effect of pretreatment of DAM (100mg/kg; im. For three days prior to administration of 47mg/kg DDVP) and DDVP on AChE activity (*in vivo*) various tissues of Rat.(Wb. Whole brain; Liv. Liver; Kid. Kidney. Tes. testis; ser. Serum) *Significance difference between treated and control groups * $P < 0.05$ and ** $P < 0.01$.