

## Effects of salt stress on ion accumulation in root and shoot of *Alhagi persarum*

Behrouz Rasouli<sup>1\*</sup>, Bahram Amiri<sup>2</sup>,

<sup>1\*</sup> - Department of Agriculture, Islamic Azad University, Rasht Branch, Iran

Email: rasouli@iaurasht.ac.ir, behrouzras@gmail.com

<sup>2</sup> - Agriculture Department, Firoozabad Branch, Islamic Azad University, Firoozabad, Iran

Email: bchamiri@iauf.ac.ir,

**Abstract.** Today, salinity has become the most important challenge for agriculture. To understand aspects of the salt tolerance of halophyte plants is important to help solve the problem of salinity in agricultural and horticultural soils. In this study we try to understand the role of root and shoot of *Alhagi persarum* in ion accumulation under salinity stress. Seeds of *A. persarum* were planted and salinity treatments (0 (control), 100, 200, 300, 400, 500 mM of NaCl and Na<sub>2</sub>SO<sub>4</sub> separately) were applied in 45 days. Na<sup>+</sup>, K<sup>+</sup>, Mg<sup>2+</sup> and Ca<sup>2+</sup> were measured in root and shoot. ANOVA results showed that salt type did not affect significantly ion content in plant, but accumulation of ions in different plant parts, except for Na<sup>+</sup>, showed significant differences. Duncan test showed that Ca<sup>2+</sup> accumulated in the root of *A. persarum* more than in the shoot. But, Mg<sup>2+</sup> and K<sup>+</sup> were higher in the shoot. Na<sup>+</sup>/K<sup>+</sup> and Na<sup>+</sup>/Ca<sup>2+</sup> ratios increased with salinity increasing. In summary, *A. persarum* tolerates salinity with the distribution of different ions in the root and shoot.

**Key words:** salinity, root, shoot, ion content, *Alhagi persarum*

### 1. Introduction

Mechanisms of plants to tolerate salinity have been intensively studied in the last decades. Most of these studies have been focused on ion content, water relations, osmolyte synthesis such as proline, betaine, carbohydrates and changes of growth indices in plants (Bohnert et al. 1995; Cicek and Cakırlar, 2008; Duan et al. 2007; Gama et al. 2007). Scientists believe that plants tolerate salinity with increasing Na<sup>+</sup> and Cl<sup>-</sup>, decreasing K<sup>+</sup>, Mg<sup>2+</sup>, Ca<sup>2+</sup> and P<sup>+</sup>, accumulate osmolytes in different organs, increasing Osmotic Potential and Water Use Efficiency and changes in Relative Water Capacity (Song et al. 2005; Zobayed et al. 2006; Gramer, 2002). One of the other things that is well known and which is one of the enigmas in salinity research, is that, although the first organ of the plant that is exposed to salinity is the root system. However, we have very little data about the role of each part of the plant in salinity tolerance (Khan et al. 2006). In fact, root, shoot and leaf have different responsibilities and operate particularly. In this study we try to understand the role of root and shoot of *Alhagi persarum* in ion accumulation under salinity stress.

### 2. Materials and Methods

*Alhagi persarum* Boiss & Buhse (camel thorn) is a valuable semi shrub and widely adapted to diverse environments in Iran. Seeds of *A. persarum* were collected and planted in plastic pots with the silica sand bed. Plants were nitrified with Hoagland's nutrient solution for 6 months (Hoagland and Arnon, 1950). Then, salinity treatment consisted of 0 (control), 100, 200, 300, 400, 500 mM of NaCl and Na<sub>2</sub>SO<sub>4</sub> separately, were applied in 45 days. Na<sup>+</sup>, K<sup>+</sup>, Mg<sup>2+</sup> and Ca<sup>2+</sup> were analyzed using an inductively coupled

plasma atomic emission spectrometer (ICP) (Navarro et al. 2006) in root and shoot of plant. Ratio of ions and shoot/root calculated. A one way ANOVA with factorial scheme was carried out to determine

<sup>1\*</sup> Corresponding author. Tel.: + (0131-3463035); fax: + (0131-3462255).  
E-mail address: (rasouli@iaurasht.ac.ir, behrouzras@gmail.com).

differences among treatments with SPSS 17.0. Significant differences between means were determined using Duncan test.

### 3. Results

A one way ANOVA showed that salt type did not affected significantly ion content and shoot/root ratio in plant, except of  $K^+$ . But accumulation of ions in different plant part, except of  $Na^+$ , show significant difference. Salt concentration had significant effect at all studied factors (table1).

Table1. ANOVA Results of ion changes in *A. persarum* organs

Dependent Variable		$Ca^{2+}$	$Mg^{2+}$	$K^+$	$Na^+$
Sources					
Salt type		0.057 <sup>ns</sup>	0.496 <sup>ns</sup>	8.351 <sup>**</sup>	0.049 <sup>ns</sup>
Plant part		376.47 <sup>**</sup>	37.56 <sup>**</sup>	389.38 <sup>**</sup>	3.314 <sup>ns</sup>
Salt concentration		5.203 <sup>**</sup>	10.43 <sup>**</sup>	25.314 <sup>**</sup>	19.22 <sup>**</sup>
Dependent Variable		$Ca^{2+}$ (shoot/root)	$Mg^{2+}$ (shoot/root)	$K^+$ (shoot/root)	$Na^+$ (shoot/root)
Sources					
Salt type		0.332 <sup>ns</sup>	0.121 <sup>ns</sup>	0.2 <sup>ns</sup>	0.202 <sup>ns</sup>
Salt concentration		4.37 <sup>**</sup>	3.4 <sup>*</sup>	5.38 <sup>**</sup>	14.17 <sup>**</sup>
Dependent Variable		$Na^+/K^+$	$Mg^{2+}/Ca^{2+}$	$Ca^{2+}/K^+$	$Na^+/Ca^{2+}$
Sources					
Salt type		0.657 <sup>ns</sup>	0.138 <sup>ns</sup>	0.413 <sup>ns</sup>	1.2 <sup>ns</sup>
Plant part		89.46 <sup>**</sup>	604.3 <sup>**</sup>	82.31 <sup>**</sup>	275.4 <sup>**</sup>
Salt concentration		18.04 <sup>**</sup>	3.45 <sup>*</sup>	3.13 <sup>*</sup>	14.44 <sup>**</sup>

• Numbers shows F value , \*  $p < 0.05$ , \*\*  $p < 0.01$ , <sup>ns</sup> non significant

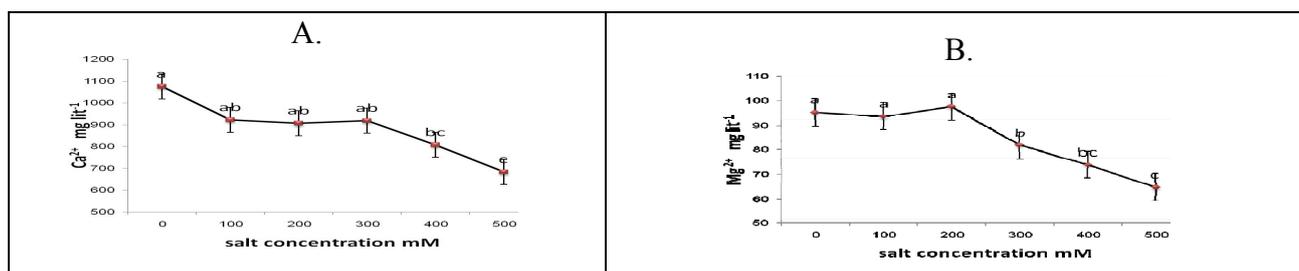
Comparison of ions at different plant organs with Duncan test showed that the root contained more Ca than shoot. But, Mg and K were more in shoot. Also, K was more at  $Na_2SO_4$  (table 2 and figure 3).

Table 2. Results of Duncan test in different plant organs and salt type

Dependent Variable	Plant part	Mean	Salt type	Mean	Dependent Variable	Plant part	Mean	Salt type	Mean
$Ca^{2+}$	shoot	430.74 b <sup>*</sup>	NaCl	891.632 a	$K^+$	shoot	533.42 a	NaCl	364.735 b
	root	1341 a	$Na_2SO_4$	880.412 a		root	239.14 b	$Na_2SO_4$	407.832 a
$Mg^{2+}$	shoot	94.687 a	NaCl	83.247 a	$Na^+$	shoot	960.34 a	NaCl	905.204 a
	root	74.167 b	$Na_2SO_4$	85.607 a		root	862.06 a	$Na_2SO_4$	917.208 a

\*: Letters indicate significant differences between treatments

Duncan test indicated that  $Mg^{2+}$ ,  $Ca^{2+}$  and  $K^+$  contents decreased with salinity increasing at all treatments (fig. 1-A,B,C and fig. 3-A,B,C). But, the concentration of  $Na^+$  ions of *A. persarum* was also influenced by salinity treatments, with seedlings exposed to greater salinity generally displaying a greater concentration of  $Na^+$  in the plant.  $Na^+$  content increased in up to 200-300 mM. But it decreased with a further increase in salinity (fig. 1-D and fig. 3-D). Fig. 2 shows that shoot/root ratio of  $Mg^{2+}$ ,  $K^+$  and  $Na^+$  decreased in up to 100 mM and increased with more increasing salt concentration. Figure 2.A indicates that  $Ca^{2+}$  decreased in shoot less than root in up to 200 mM and then it became inverse. Also,  $Ca^{2+}$  content in shoot is less than root at all salt concentrations whilst  $Mg^{2+}$ ,  $K^+$  and  $Na^+$  in shoot are more than root (fig. 2).



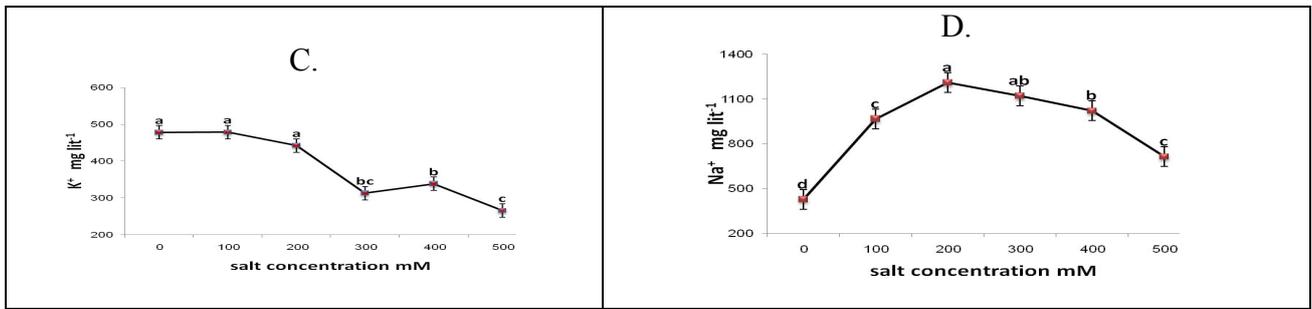


Fig. 1: Duncan test of K<sup>+</sup>, Mg<sup>2+</sup>, Ca<sup>2+</sup> and Na<sup>+</sup> at different salinity concentrations. Bars represent mean ± standard error. Different letters represent a significant difference P < 0.05 between treatments.

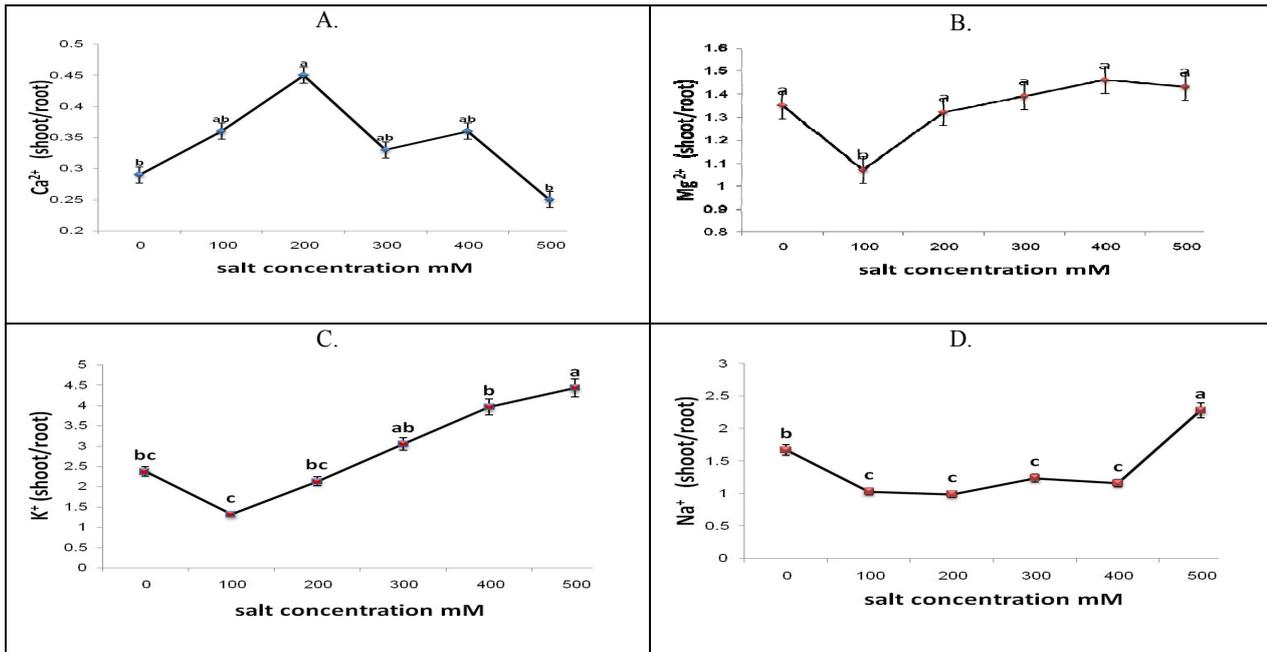
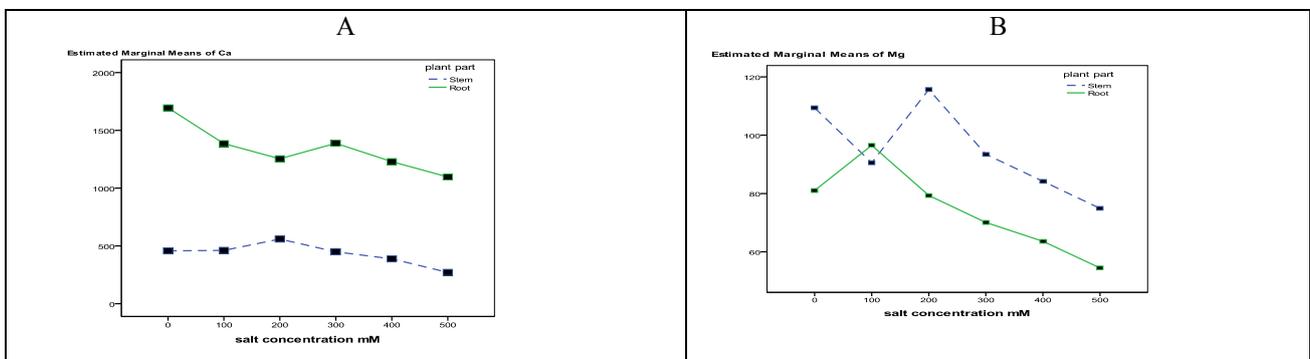


Fig. 2: Duncan test of shoot/root ions ratio at different salinity concentrations. Bars represent mean ± standard error. Different letters represent a significant difference P < 0.05 between treatments.

Moreover, Figure 3 indicated that changes trend was not coincided in root and shoot, especially at low salinity. The root content of K<sup>+</sup> and Mg<sup>2+</sup> tended to increase, while its shoot content tended to decrease. There was relatively small reduction in stem Ca<sup>2+</sup> in comparison to the very large reduction of root.



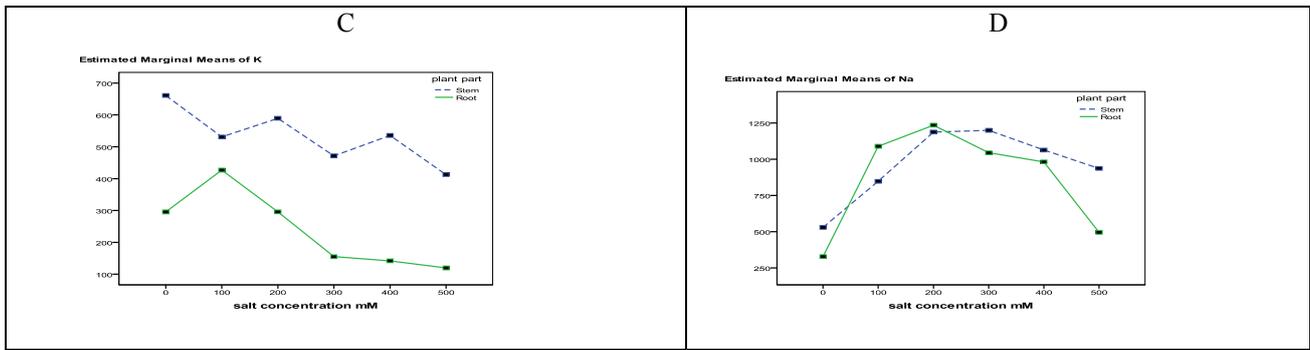


Fig. 3: estimated marginal means of ions at different salt concentration and plant parts.

Figure 4.A and B indicate that  $\text{Na}^+/\text{K}^+$  and  $\text{Na}^+/\text{Ca}^{2+}$  ratio increased with salinity increasing in shoot and root of *A. persarum*. But they decreased at high salinity in root.  $\text{Na}^+$  increasing simultaneous with  $\text{K}^+$  and  $\text{Ca}^{2+}$  decreasing. In addition,  $\text{Ca}^{2+}/\text{K}^+$  ratio decreased at low salinity in root and then increased.  $\text{Ca}^{2+}$  and  $\text{K}^+$  changed in shoot, similarly and  $\text{Ca}^{2+}/\text{K}^+$  ratio did not vary significantly in shoot with salinity increasing.

$\text{Mg}^{2+}/\text{Ca}^{2+}$  ratio decreased at moderate salinity in shoot whilst it increased in root. It did not vary significantly at control and high salinity in root and shoot.

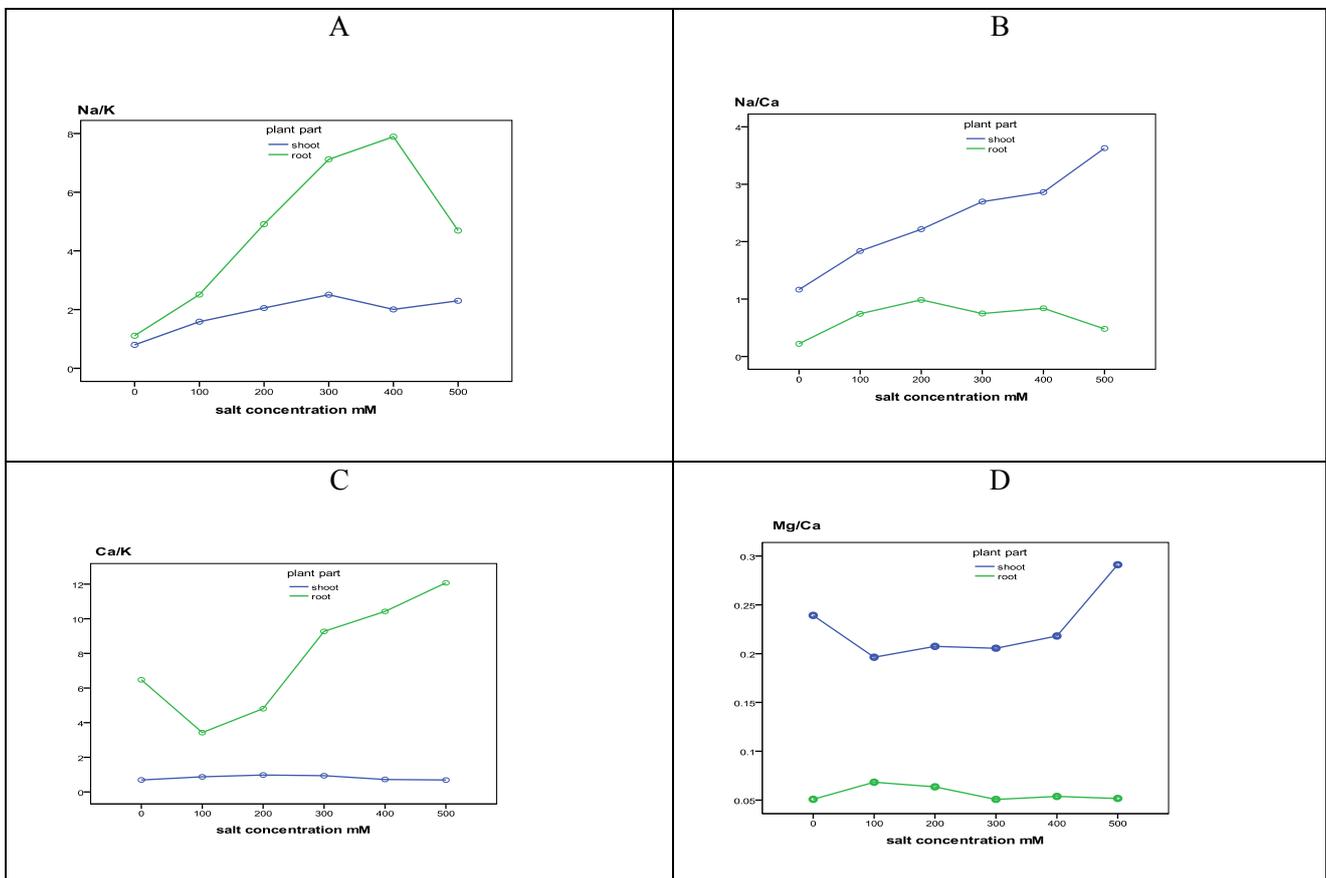


Fig. 4: ions ratio changes at different salt concentrations and in different plant organs.

#### 4. Conclusion

According to our results, ions accumulation in *A. persarum* was influenced by salt concentrations and plant organs, whilst salt type did not affect ion content. *A. persarum* uptakes  $\text{Na}^+$  in saline environments and repulses  $\text{K}^+$ ,  $\text{Ca}^{2+}$ ,  $\text{Mg}^{2+}$ . El-Fouly *et al.* (2004) in tomato, Kurban (2005) in *alhagi pseudoalhagi* and Arndt *et al.* (2004) found that  $\text{Na}^+$  and  $\text{Cl}^-$  uptake markedly increased with increasing NaCl growth medium. Khan *et al.* (2006) reported that  $\text{K}^+$  and  $\text{Mg}^{2+}$  decreased with salinity increasing in alfa alfa. This study showed that  $\text{Ca}^{2+}$  decreased in shoot more than root at high salinity. Cramer *et al.* (1990) and Khan *et al.* (1998) indicated that decrease of  $\text{Ca}^{2+}$  in aboveground organs is an important indicator of salinity stress.

Jie et al. (2008) indicated that the halophyte plants accumulated more  $\text{Na}^+$  and  $\text{Cl}^-$  in aboveground tissues compared to roots. Kurkova et al. (2002), also, showed that the contents of  $\text{Na}^+$  and  $\text{K}^+$  increased in the root tissues at low salinity and the contents of sucrose, proline, and betaines decreased as compared. They said that halophyte plants are capable of withstanding abrupt salt stress, due to the shift of osmoregulatory systems (ionic and organic) and isolation of the excess of salinating ions by their accumulation in the central vacuole through pinocytosis, as well as in small cytoplasmic vacuoles. Therefore, it can be suggested that pinocytosis and small cytoplasmic vacuoles were extended in shoots more than roots, when the plant exposed to salinity. Increasing of  $\text{Na}^+/\text{K}^+$  and  $\text{Na}^+/\text{Ca}^{2+}$  ratio can be related to selective absorption of cell membrane and salinity tolerance. It has been reported by Gorham (1997).  $\text{Ca}^{2+}/\text{K}^+$  and  $\text{Mg}^{2+}/\text{Ca}^+$  ratio indicated that there is competition among  $\text{Ca}^{2+}$ ,  $\text{Mg}^{2+}$  and  $\text{K}^+$  absorption in plant. Also,  $\text{Ca}^{2+}/\text{K}^+$  ratio in shoot and  $\text{Mg}^{2+}/\text{Ca}^{2+}$  ratio in root of *A. persarum* show cooperation relation between them. These agree with Belda and Ho (1993) and Robert and Susan (2004). In general, the most changes of ions in different plant organs of *A. persarum* and salt concentration occur at 100-300 mM of salinity. It confirms that it is a moderate halophyte.

## 5. Refrences

- [1] Arndt S. K., C. Arampatsis, A. Foetzki, X. Li, F. Zeng, X. Zhang, .2004. Contrasting patterns of leaf solute accumulation and salt adaptation in four phreatophytic desert plants in a hyperarid desert with saline groundwater. *J. Arid Environments* 59 259–270.
- [2] Belda R. M., and Ho. L.C. 1993. Salinity effects on network of vascular bundles during tomato fruit development. *J. Hort. Sci.* 68:557-564.
- [3] Bohnert H.J, Nelson D.E, and Jensen R.G. 1995. Adaptation to environmental stresses. *Plant Cell* 7:1099-1111.
- [4] Cicek N. and H.Cakırlar. 2008. Effects of salt stress on some physiological and photosynthetic parameters at three different temperatures in six Soya Bean Cultivars. *J. Agronomy and Crop Science*,194(1)34-46.
- [5] Colmer, T.D., E. Epstein, and J. Dvorak. 1995. Differential solute regulation in leaf blades of various ages in salt-sensitive wheat and a salt-tolerant wheat 3 *Lophopyrum elongatum* .Host. A. Lo<sup>o</sup> ve Amphiploid. *Plant Physiol.* 108:1715–1724.
- [6] Cramer G.R, Lyuchli A, and Polito V.S.1990. Displacement of  $\text{Ca}^{2+}$  by  $\text{Na}^+$  from the plasmalemma of root cells. response to salt stress. *J.Plant Physiol.*79:207- 277.
- [7] Duan. D.Y, Wei, Q. L, Xiao, J. L, Hua. O. and Piny, A. 2007. Seed germination and seeding growth of suaeda salsa under salt strees, *Bot. Fennici.* Vol. 44- 161-169.
- [8] El-Fouly, M.M., Z.M. Moubarak and Z.A. Salama. 2002. Micronutrient foliar application increases salt tolerance of tomato seedlings. *Proc. Inter. Symp. On “Techniques to Control Salination for Horticultural Productivity”* Eds. U. Akosy *et al.*, *Acta Hort.* No. 573: 377-385.
- [9] Gama, P, S., Inanaga, S., Tanaka, K. and Nakazawa, R. 2007. , Physiological response of common bean .*Phaseolus vulgaris* L. seedlings to salinity stress, *African J. Biotechnology*, 6( 2) pp.79-88 ,
- [10] Gramer, G. R., 2002, Uptake and role of ions in salt tolerance. In: Jaiwal PK, Singh RP, Gulati A .eds. strategies for improving salt tolerance in higher plants. Oxford and IBH Pupliching, New Delhi, pp: 55-85.
- [11] Gorham, J.1997. Mechanism of salt tolerance of halophytes. In: Choukr Allah, R. Malcom. C. V., Hamdy, A., eds. *Halophytes and biosalin agriculture.* New York: Marcel Dekker, Inc., 31-53.
- [12] Hoagland D.R. and Arnon D.I. 1950. The water-culture method for growing plants without soil. *Calif Agric Exp Stn Circ* 347:1-39.
- [13] Jie Z, Z. FanJiang, Arndt S. K., .2008. Growth, physiological characteristics and ion distribu-tion of NaCl stressed *Alhagi sparsifolia* seedlings *Chinese Science Bulletin* 2008, 53.zkII. 169-176, ISSN: 1001-6538
- [14] Khan, M.A, Ansari, R, Gul M. 2006. Crop diversification through halophyte production on salt prone land resources. *CAB Reviews: Perspective in Agriculture, Veterinary Science, Nutrition and Natural Resources*.1, No 048,8

- [15] Khan M.A, Ungar I.A, Showalter A.M.1998. NaCl-induced accumulation of glycinebetaine in four subtropical halophytes of Pakistan. *Physiol Plant*. 102:487-492.
- [16] Kurban H, Saneoka H, Nehira K, Adilla R .1999. Effect of Salinity on Growth, Photosynthesis in Leguminous Plant *Alhagi pseudoalhagi* .*Bieb. J. of Soil science and plant nutrition*
- [17] Kurkova E. B., G. Kalinkina, O. K. Baburina, N. A. Myasoedov, and T. G. Naumova. 2002. Responses of *Seidlitzia rosmarinus* to Salt Stress, *Biology Bulletin*, vol. 29, No. 3, 221-228
- [18] Navarro A., S. Banon and E. Olmos. 2006. Effects of sodium chloride on water potential components, hydraulic conductivity, gas exchange and leaf ultrastructure, *Plant Science*. 172 .p:473–480.
- [19] Robert W. P and Susan L. U. 2004. Effects of salinity on growth and photosynthesis of three California tidal marsh species, *Cell Biol. Intern*. 17:839-845
- [20] Song, J. Zhao, K.F. Fan, H. Sun, M.X. Wang B.Z. Zhang, S.Q. and Ungar, I.A. 2005, Two Na<sup>+</sup> and Cl<sup>-</sup> hyperaccumulators of the Chenopodiaceae. *J.Integrative Plant Biol*. 47, 311-318
- [21] Zobayed .M.A. Murch, S.J. El-Demerdash, M.A. 2006. Nacl enhances growth and morphogenesis potential of *Alhagi graecorum*. *J in vitro cellular and development biology plant*.42, 607-613
- [22] Edward, P. G.; Brown, J. J. Salt tolerance and crop potential of halophytes. *Plant Science, Berlin*, v.138, n.2, p.227-253, 1999.